
प्रयोगशाला के जानवरों के प्रजनन,
देखभाल, प्रबंधन और आवास के लिए कोड
भाग 2 प्रयोगशाला खरगोश
(दूसरा पुनरीक्षण)

**Code for Breeding, Care,
Management and Housing of
Laboratory Animals**

Part 2 Laboratory Rabbits

(Second Revision)

ICS 65.020.30

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FOREWORD

This Indian Standard (Part 2) (Second Revision) was adopted by the Bureau of Indian Standards, after the draft finalized by the Animal Husbandry and Equipment Sectional Committee had been approved by the Food and Agriculture Division Council.

Laboratory animals bred and maintained scientifically are necessary to get comparable results in biological experiments. The present standard lays down guidelines for breeding, care, management and housing of laboratory rabbits so that large number of animals of uniform quality are available. It is hoped that this would streamline and step-up facilities for pharmaceutical, pesticidal efficacy and biologically oriented research. Laboratory are used extensively for biological research and in biomedical field. This standard is published in 10 parts, these parts are namely:

- Part 1 Laboratory mice and rats
- Part 3 Laboratory guinea — Pigs
- Part 4 Laboratory golden hamsters
- Part 5 Laboratory snakes
- Part 6 Laboratory cotton rats (*Sigmodon Hispidus* and *Sigmodon Hispidus Hispidus*)
- Part 7 Laboratory frogs
- Part 8 Laboratory chicks
- Part 9 Laboratory pigeons
- Part 10 Laboratory mosquitoes

This standard was first published in 1970 and first revised in 1979 for incorporating modifications in cage size, maximum population per cage, and areas for different sections of laboratory animal facility. In this revision, the standard has been comprehensively reviewed and aligned with the *Breeding of and Experiments on Animals (Control and Supervision) Rules*, 1998 under *Prevention of Cruelty to Animals Act*, 1960 and also with 'CPCSEA guidelines for laboratory animal facility, 2015'. In this revision following major modifications have been done:

- a) Details of disease control procedures for laboratory rabbits to be used;
- b) Types of records to be kept; and
- c) Conduct of painful and invasive procedures and humane ends.

The composition of the Committee responsible for the formulation of this standard is given in [Annex A](#).

For the purpose of deciding whether a particular requirement of this standard is complied with, the final value observed or calculated, expressing the result of a test or analysis, shall be rounded off in accordance with IS 2 : 2022 'Rules for rounding off numerical values (*second revision*)'. The number of significant places retained in the rounded off value should be the same as that of the specified value in this standard.

*Indian Standard***CODE FOR BREEDING, CARE, MANAGEMENT AND
HOUSING OF LABORATORY ANIMALS****PART 2 LABORATORY RABBITS***(Second Revision)***1 SCOPE**

This standard (Part 2) prescribes optimum conditions for housing, sanitation, personnel hygiene, feeding, watering, disease control, etc, about care, breeding, and management of laboratory rabbit.

2 TERMINOLOGY

Following terminology shall be used for the purpose of this standard:

2.1 Laboratory Animal Facility — A place where laboratory animals are scientifically reared or kept for breeding, maintenance, or experimentation.

3 LOCATION AND HOUSING

3.1 Laboratory animal facilities should be located in dry and well-drained areas free from smoke, obnoxious fumes, dust, noise, and extremes of temperatures.

3.2 Laboratory animal facilities should have secured access for the animal users, uninterrupted water and electricity supply and it should be away from residential areas and have a good source of fresh air.

3.3 Careful planning should be done to place laboratory animal facility areas near research laboratories but separated from them by barriers, such as entry locks, corridors, or floors.

3.4 Laboratory animal facility should have the following areas:

- a) Animal rooms for breeding, holding experimentation, quarantine, isolation, etc;
- b) Service areas for cage sanitation, sterilization, washing, stores (feed, bedding, cages, and general facility storage);
- c) Space for waste handling and storage including refrigerated/cold storage for dead animal wastes before disposal and areas for service corridors to facilitate the movements of equipment and personnel;

- d) Disease diagnostic laboratory, procedure room, necropsy room.
- e) Office and support areas for staff and others; and
- f) Mechanical, electrical, heating, ventilation and air-conditioning (HVAC), building monitoring system (BMS), etc.

3.5 The area for each category depends on the number of animals, species housed, and the type of facility (conventional or barrier).

3.6 There should be adequate separation between clean and dirty areas/operations to avoid cross-contamination. A 'dirty' operation should not be proximate to 'clean' areas.

4 CONSTRUCTIONAL REQUIREMENTS**4.1 Walls**

The walls should be fire-resistant, vermin-proof, moisture-resistant, and easily cleanable, with high-build coating (HBC) or solid panels. Walls in high-altitude areas should be thermally insulated.

4.2 Interior

The inner surface of the walls should be waterproof, smooth, and easy to clean.

4.3 Roof

A concrete roof with adequate insulation is preferable. If the roof is constructed of sheets or tiles, a false ceiling with moisture-resistant gypsum board, latex paint or HBC, and sealant at the intersection should be used to achieve air and water tightness.

4.4 Floor

4.4.1 The floor shall be constructed of concrete base with monolithic or having a minimal number of joints.

4.4.2 The floor should be relatively smooth for easy cleaning, skid-proof, and waterproof.

4.4.3 The floor should be resistant to wear and tear, disinfectant wash, high-temperature water, acids, and solvents.

4.4.4 The floor should be free from cracks and crevices. The junctions of the floor, ceiling, and walls of the rooms should be coved to avoid the accumulation of dirt and dust and for effective cleaning and disinfection.

4.4.5 The floor should be capable of withstanding 6 kPa (125.3 psf) live load to dampen vibration.

4.4.6 Vitrified tiles, seamless epoxy, or resinous floor coatings are preferred.

4.4.7 In the washing and sanitation area, ceramic tiles with non-slippery flooring shall be utilized.

4.5 Doors

4.5.1 The doors shall be designed with a seamless surface and frame, be unable to carry bacteria, and will not warp when in contact with disinfectants.

4.5.2 The door should have automatic door closure mechanisms along with shielded handles, door seals, kick plates, jamb guards, etc.

4.5.3 Doors should be opened inwards towards animals.

4.5.4 The door should also have a viewing panel for observation and safety reasons.

4.5.5 Adequate latches and locking arrangements should be provided.

4.5.6 Doors should be large enough to allow the easy passage of racks and equipment.

4.5.7 Doors should fit tightly within their frames and door seals should be placed to control air movement and to prevent vermin entry.

4.6 Windows

Exterior windows inside the rabbit rooms are not advised, as they will affect the photoperiod of rabbits.

4.7 Openings and Exhaust

4.7.1 Each animal room should have adequate air-exhaust provisions and the openings should be vermin-proof and sealed.

4.7.2 There should be a balanced exhaust air system relative to the air handling unit (AHU) supply.

4.7.3 Exhaust air should be cleared outdoors without recirculation inside the room.

4.7.4 There should be a separate exhaust system for the quarantine/isolation room, sanitation and washing area, and necropsy rooms.

4.8 Corridors

4.8.1 Corridors should have adequate space for the movement of personnel and equipment inside the room.

4.8.2 Corners between floor and walls should be coved for easy and thorough cleaning.

4.9 Floor Drains

Floor drains and washbasin are not recommended in rodent rooms.

4.10 Airlock for Main Entrance

4.10.1 It is desirable to have an airlock followed by an air shower after wearing the necessary personal protective equipment (PPE) at the main entrance of the animal facility.

4.10.2 Change area and air shower facilities should be located near the personnel entrance to the facility.

4.11 Light

4.11.1 There should be adequate arrangements for lighting inside the rooms.

4.11.2 Standard fluorescent light or cool white LED light is preferred with an automatic timer system to maintain the photo period of 12 h/12 h to 10 h/14 h light/dark with illumination less than 400 lux and preferably about 325 lux unit when measured approximately 1 m above the floor.

4.11.3 The lighting should be diffused throughout the animal room and fixed accordingly so that the bottom-most cages in the animal rooms also receive similar light intensity.

4.11.4 Lighting fixtures, switches, etc, shall be designed and placed accordingly to prevent vermin/insects' entry.

4.11.5 All the control switches should be kept, as far as possible, outside the rooms.

4.12 Ventilation and Temperature

4.12.1 An ideal temperature of 20 °C to 26 °C (depending on the species/strain used and lactating

mother and pups) with 40 percent to 70 percent relative humidity should be maintained.

4.12.2 A 10 to 15 air change/hour with 100 percent fresh air is recommended in animal rooms. However, consideration should be given to possible heat loads and the number of animals involved.

4.12.3 The heating, ventilation, and air conditioning (HVAC) system of each room should control the temperature and relative humidity through the building monitoring system (BMS).

4.12.3.1 In air-conditioned rooms, each animal room should have temperature and relative humidity gauges mounted on the outside wall.

4.12.3.2 When employing a room air conditioner system, necessary provisions should be provided for controlling temperature, relative humidity, and airflow.

4.12.3.3 The controls for lighting, air-conditioning, exhaust fans, etc, should also be located outside the animal room.

4.12.3.4 Exhausts should be installed close to ground level when cages are placed parallel to walls.

4.12.3.5 Racks should be arranged accordingly in a room to optimize air exchange.

4.12.3.6 Ammonia content in an animal room shall not exceed 10 ppm and intra-cage ammonia levels should be kept to a minimum or absent.

4.12.3.7 Alternate or emergency power should be available in the event of power failure to maintain critical services like HVAC, lighting, and housing equipment in animal rooms and other essential areas.

4.13 Vermin Control

4.13.1 All interior animal housing and support units must ensure that windows, doors, and exterior walls should be sealed to prevent the entrance of pests and predators. Interior walls, drains, and vents shall be checked for cracks and leaks and repaired as needed.

4.13.2 Effective vermin management procedures should be in place to ensure that the livestock facility is free of pest and rodent infestation.

4.14 Noise and Vibrations

It is recommended to control the noise and vibration in animal rooms to preferably less than 85 db.

5 HOUSING AND EQUIPMENT

5.1 Rack

Racks may be floor-mounted, wall-mounted, or on casters. If racks are mobile, a mechanism to secure the rack should be included as the motion of the animals within the cage can cause rack motion.

5.2 Cages

5.2.1 The cages should be made of smooth corrosion-resistant material that does not tear off or get gnawed away by the animal and preferably have rounded corners and bottoms made of a suitable non-corrosive material. It should have provision for suspended cages (4 to 6) as per the recommended space requirement and equipped with, pans, feeders, watering bottle and nesting material.

5.2.2 It is recommended to use cages made of material such as polypropylene (opaque) or polycarbonate, polysulfone, and polyetherimide (transparent), with wire mesh tops or any other similar suitable material that is easy to clean and resistant to sterilization at 121 °C and chemical treatment.

5.2.3 The lid should provide a space for keeping the water bottle.

5.2.4 Wire lids for cages should be carefully selected to prevent toe injuries.

5.2.5 Each cage should be provided with proper enrichment and nesting material.

5.2.6 The cages should be impervious to liquids and moisture, easily cleaned and sanitized.

5.2.7 Static isolator cages must be cleaned once a week to avoid excessive ammonia and carbon dioxide levels.

5.2.8 The cage should meet the minimum space requirements for the rabbit and the caging must be without sharp edges or protrusions which may cause injury.

5.2.9 A proper paper liner should be placed beneath the rabbit cages for easy cleaning.

5.2.10 The cage should have provisions for feed water, and waste disposal and it should be made up of good quality material usually stainless steel

5.2.12 The slots or openings for fecal passage should be approximately 0.5 inches to 0.7 inches.

Flooring configuration can be either slatted, tubular, wire grid, or plastic with perforations.

5.2.13 The minimum floor area recommended for rabbits in cages (based on their weight/size) and interior cage height shall be as given in [Table 1A](#). The minimum floor area recommended for female rabbits with litters in cages (based on their weight/size) and interior cage height shall be as given in [Table 1B](#).

Table 1A Space Requirements for Rabbits

(Clause [5.2.13](#))

Sl No.	Body Weight (kg)	Floor Area/ Rabbit Feet ² (m ²)	Interior Cage Height Inches (cm)
(1)	(2)	(3)	(4)
i)	> 2	1.5 (0.14)	14 (35.56)
ii)	2 to 4	3.0 (0.28)	14 (35.56)
iii)	4 to 5.4	4.0 (0.37)	14 (35.56)
iv)	< 5.4	5.0 (0.46)	14 (35.56)

Table 1B Space Requirements for Females with Litters

(Clause [5.2.13](#))

Sl No.	Body Weight, kg	Floor Area/ Rabbit, Feet ² (m ²)	Interior Cage Height, Inches (cm)
(1)	(2)	(3)	(4)
i)	< 2	4.0 (0.37)	13.8 (35)
ii)	2 to 4	5.0 (0.46)	13.8 (35)
iii)	4 to 5.4	6.0 (0.56)	13.8 (35)

5.3 Water Supply

5.3.1 There should be an adequate supply of drinking water free from any type of contamination where water can be treated with reverse osmosis, acidification, or steam sterilization before it is dispensed to the animals.

5.3.2 The water bottle should be of polypropylene, polycarbonate, polysulfone, or higher quality that can withstand autoclaving.

5.3.3 Water bottles should be provisioned with suitable nozzles.

5.3.4 Water should be given *ad libitum* and periodic changing of the water bottles is necessary to prevent contamination.

5.3.5 Water should be provided by water bottles or automatic watering systems.

5.3.6 Stainless steel should be used for sipper tubes. If rubber stoppers are used, there should be a metal guard.

5.4 Feeding System

5.4.1 The feeding system should be of J-type feeder that is attached securely to the cage.

5.4.2 The rabbit's food access point should be 3 inches to 4 inches off the floor, with an opening large enough to prevent the rabbit's head from becoming trapped.

5.4.3 Necessary feed bowls or troughs can be attached to the inside of the cage.

5.4.4 Feeding devices, including bowls and feed hoppers should be attached to the cage exterior.

5.4.5 To minimize contamination by urine or feces, feed should not be placed directly on the bottom of the cage.

5.4.6 In each primary enclosure housing a female with a litter less than one month old, an appropriate nest box containing clean nesting material must be provided.

5.4.7 Nest boxes must be made of stainless steel and must be sanitized regularly.

5.5 Other Requirements

5.5.1 An elevated platform should be provided when rabbits are housed in cages

5.5.2 There should be enough area for group housing/social housing, to exercise, and should have necessary enrichment to allow the animals to execute a wide range of natural behaviours.

5.5.3 Rabbits should be housed in stable, suitable groups as soon as possible after weaning, with immature animals of the same age and sex.

5.5.4 Hay should be provided for foraging, play, nest building, and for a varied diet.

5.5.5 Rabbits should always have something to nibble on (for example, wooden blocks) for

enrichment, chin-marking, and to keep their teeth from overgrowing.

5.5.6 Rabbits should have access to substitute burrows (for example, plastic crates, sections of appropriately sized PVC pipe) for retreat in fear-provoking situations and to manage social interactions.

5.5.7 There should be good visibility out of the pen (for example, mesh or plastic wall) so that the animals can overlook their surroundings and see approaching personnel.

5.5.8 Rabbits should experience gentle and frequent handling from early in life.

5.5.9 The provision of environmental enrichment is particularly important for singly housed or caged rabbits, and the provision for adequate exercise is fundamental to the normal skeletal and muscular development and maintenance of all laboratory rabbits.

5.5.10 Suitable enrichment for rabbits includes roughage, hay blocks, or chew sticks as well as an area for withdrawal. In floor pens for group housing, visual barriers and structures to provide refuges and lookout behavior should normally be provided. For breeding nesting material and a nest box are appropriate provisions.

6 FOOD AND BEDDING

6.1 Bedding

6.1.1 Dry, non-absorbent material like fine paper cuttings, wood wool, or cotton shall be placed in the cage corner reserved for nesting before delivery.

6.1.2 The bedding shall be stored in a dry and clean place.

6.2 Feeding

6.2.1 Rabbits are fastidious eaters and may reject foods solely based on odor, texture, form, or method of presentation.

6.2.2 Rabbits may be fed ad libitum, however, to prevent obesity the amount of pelleted feed is substituted with a good quality grass hay or greens to keep rabbits from gaining excess weight and to prevent the formation of hairballs.

6.2.3 Because the rabbit's intestinal microflora is sensitive to the intestinal contents like pH, carbohydrate content, etc, food changes should be

made gradually, especially for 4 to 12 week old rabbits.

6.2.4 The new food should be gradually added, and the old food decreased over a 4 to 5 day period to allow the flora to adjust.

6.2.5 Under certain conditions, rapid food changes can result in the proliferation of *Clostridial* and *Coliform* bacteria, which may result in the death of the animal.

6.2.6 Vitamin supplementation is normally not necessary for rabbits on an alfalfa-based diet.

6.2.7 Rabbits have a high-water turnover compared to other species of animals and consume twice as much water as food on a per-weight basis. The average water consumption is 120 ml/kg body weight per day.

6.2.8 The pelleted feed should contain a proximate analysis report of each batch including date of manufacturing and expiry. The feed should be stored in room having temperature below 23 °C and relative humidity below 50 percent. The feed intake and nutritional requirement are recommended to be approximately as given in [Table 2](#) and [Table 3](#) respectively.

7 CLEANING, STERILIZATION AND WASTE DISPOSAL

7.1 Cleaning of Laboratory Animal Facility

- The laboratory animal facility should have adequate sanitizing and washing facilities;
- Suitable detergents, and disinfectants, should be used in rotation for floor cleaning; and
- Deodorants designed to mask animal odors should not be used in housing.

Table 2 Daily Feed Intake in Rabbits

(Clause [6.2.8](#))

SI No.	Age Group	Feed Intake
(1)	(2)	(3)
i)	Adult rabbits	5 g/100 g body weight
ii)	Pregnant does	175 g to 225 g
iii)	Lactating does	225 g to 450 g
iv)	3 to 6 weeks	50 g to 75 g
v)	6 to 8 weeks	125 g to 150 g

Table 3 Nutrition Requirement in Rabbits

(Clause 6.2.8)

SI No.	Nutrients	Growth	Maintenance	Gestation	Lactation
(1)	(2)	(3)	(4)	(5)	(6)
i)	Digestible energy (kcal)	2 500	2 100	2 500	2 500
ii)	Total digestible nutrients (%)	65	55	58	70
iii)	Crude fiber (%)	10 to 12	14	10 to 12	10 to 12
iv)	Fat (%)	2	2	2	2
v)	Crude protein (%)	16	12	15	17

7.2 Cleaning of Cages, Bottles, Tubes

- a) Cages should be sanitized before animals are placed in them. Generally, cages (including lids, food hoppers, etc) should be changed and sanitized at least once a week;
- b) However, a decision to change the frequency of such bedding changes or cage washing should be based on factors such as the concentration of ammonia, appearance of the cage, condition of the bedding, and number and size of the animals housed in the cage;
- c) The routine sterilization of drinking bottles and tubes should be done once a week;
- d) Suitable disinfectants based on the manufacturer's recommendation may be used for cage washing and water bottle washing; and
- e) Autoclaving of cages should preferably be done at 121 °C for 15 min at 1.05 kg/cm² (15 lb) pressure.

7.3 Waste Disposal

All wastes from the animal facility should be collected and disposed of as per the biosafety norms and categorized accordingly in respective collection colour-coded bags as classified below and disposed accordingly safely and hygienically:

a) Yellow bags

It shall handle following components:

- 1) Animal anatomical waste — Experimental animal carcasses, body parts, organs, and tissues, including the waste generated from animals used in experiments or testing in laboratory animal facility, cytotoxic drugs and antibiotics with their vials;

2) Liquid waste generated due to the use of chemicals in the production of biological and used or discarded disinfectants; and

3) Discarded beddings contaminated with blood or body fluid.

b) Red bags:

1) To handle wastes generated from disposable items such as tubing, bottles, and syringes (without needles and fixed needle syringes) vacutainers with their needles cut) and gloves.

c) White puncture-proof containers:

1) To handle needles, syringes with fixed needles, needles from needle tip cutters or burners, scalpels, blades, or any other contaminated sharp object that may cause punctures and cuts. This includes both used, discarded, and contaminated metal sharps.

d) Blue bags:

1) To handle broken or discarded and contaminated glass including medicine vials and ampoules except those contaminated with cytotoxic wastes.

7.4 Storage and Transport

a) Waste materials should be removed from the animal rooms at frequent intervals;

b) If storage of waste materials before removal is necessary, the holding area should also be located in a place physically separate from the main animal rooms and free from flies, cockroaches, rodents; and other pests;

- c) The waste materials should be finally disposed of preferably by incineration or by other methods prescribed by local municipal or civic bodies;
- d) The duration of storage should not exceed 24 h;
- e) Each bag may be clearly labeled with details of the facility;
- f) The waste should be transported for treatment either in trolleys or in a covered wheelbarrow; and
- g) Manual loading should be avoided as far as possible. The bags/containers containing biomedical wastes should be tied/lidded before transportation or as suggested by local bodies.

8 BREEDING

8.1 A nesting box of size 500 mm × 275 mm × 250 mm approximately with bedding material such as straw or clean, shredded paper should be provided before the expected kindling date.

8.2 The nest box should not be placed in the corner of the cage or pen where the doe typically urinates.

8.3 Entire male rabbits should be separated from other males at sexual maturity (12 to 14 weeks) and housed individually with visual and olfactory contact with other rabbits.

8.4 Rabbit reproductive biology is given in [Table 4](#) for general guidance.

9 PERSONNEL

9.1 All persons recruited for work in laboratory animal facility should have a natural aptitude for handling animals and they should be given training for their particular duties before they are given independent charge of the work.

9.2 The persons required to handle animals should be of sound health and should not be suffering from infectious diseases communicable to animals or other fellow workers. Medical examination before recruitment and at periodical intervals should be arranged.

9.3 The workers before entering the animal premises, should wash their hands, feet, and face with soap and water. They should take a shower where necessary.

9.4 He/she should either completely change the street clothes with uniforms or apparel issued specifically for the purpose or at least cover his/her clothes, provided they are reasonably clean, with protective apparel. The protective apparel should consist of a cotton apron or coat that covers the body from neck to knees and arms up to elbows, hand gloves, a suitable cover for the head, a white cloth mask that covers the nose and mouth, and soft shoes or footwear or disposable shoe cover. These articles should be issued to individuals by name and should be stored when not in use in lockers or shelves where they shall not be contaminated with the clothes of others.

Table 4 Rabbit Reproductive Biology

(Clause 8.4)

SI No.	Particular	Age and Period
(1)	(2)	(3)
i)	Puberty usually	5 to 7 months
ii)	Breeding age	1 to 3 years (some remain productive for up to 5 or 6 years)
iii)	Breeding life	Average of 1 to 3 years, although
iv)	Ovulation	10 to 13 hours after copulation
v)	Gestation length	31 to 35 days
vi)	Litter size	7 to 9
vii)	Weight of kids (g)	30 to 100
viii)	Weaning age	5 to 8 weeks
ix)	Ears open	8 days
x)	Eyes open	9 to 11 days

9.5 Before the workers leave the laboratory animal facility for the day, their apparel should be removed and placed in their proper place. In no case should the clothes, shoes, etc, be allowed to be taken out of the premises. Clothing should be laundered every day.

9.6 During work time, workers should be instructed to wash their hands with soap and water as often as necessary. Individual absorbent towels or other hygienic facilities for drying hands should be available near the wash basins.

9.7 Persons should be allotted work in such a manner that the same person handles the same batch of animals daily except in emergencies.

9.8 Access to the animal housing facilities by unauthorized personnel should be restricted.

9.9 No material other than those required for work should be permitted to be taken into the animal rooms.

9.10 Eating food, chewing pan, or smoking in the rooms should be prohibited.

9.11 Toilets for workers should be located outside the animal rooms and every time they visit the toilet rooms, they should follow all the procedures required while leaving or entering the laboratory animal facility.

9.12 Floor mats soaked in a suitable disinfectant may be placed at the entrance of each block of animal rooms/clean area so that the footwear may be wiped against them before entering or leaving the rooms.

9.13 There should be adequate contingency plans to cover such emergencies as flooding and fire, or the breakdown of lighting, heating, cooling, or ventilation.

9.14 Occupational Health Program

The occupational health program is voluntary but it should be highly encouraged for all members of any institute who work in laboratory animal facilities or have substantial animal contact. This includes animal resource personnel, research technicians, research investigators, faculty, and staff. The program consists of the following if deemed necessary by the occupational health physician.

9.15 Completion of Occupational Health Questionnaire

9.15.1 Review of the health questionnaire and risk assessment performed by the occupational health

physician who will determine which test will be performed on the individual.

9.15.2 These procedures shall be offered at no cost to the employee/staff and shall be conducted under the direction of a licensed healthcare professional.

9.15.3 Vaccination/medical records should be maintained in the employee's personnel folder and shall be provided upon written request for copying to the subject employee or anyone having written consent of the subject employee.

10 DISEASE CONTROL PROCEDURES

10.1 Adequate precautions, including quarantine, periodical inspection, and preventive measures, should be enforced to prevent infection of the animals by ectoparasites or infectious diseases.

10.2 All fresh batches of animals received should be quarantined for at least one week (preferably three weeks) in separate premises before being introduced into the regular stock.

10.3 Only healthy animals should be allowed to enter the premises.

10.4 Infections, if any are noticed, should be immediately eliminated by:

- a) Euthanizing the infected animals, if possible; and
- b) Locating and treating the source of infection.

10.5 All dead animals should be immediately removed and disposed off.

10.6 If any mortality occurs in the colony, the cause of death should be investigated and if traced, steps should be taken to prevent the spread of the infection.

10.7 Well-equipped disease diagnostic laboratory facilities should be provided in each animal facility.

10.8 Morbid/sick animals should be sent from breeding rooms for diagnosis and thereafter disposed of.

10.9 In the case of very small animal facilities, collaboration with competent laboratories may be arranged for such investigation.

10.10 Anesthesia and Euthanasia

10.10.1 The principles and guidelines for anesthesia and euthanasia should be followed as recommended by CCSEA guidelines for anesthesia and euthanasia.

10.10.2 Procurement of anesthetic drugs and their custody and stocking and administration shall be done by a qualified veterinarian only.

10.11 Health Monitoring

10.11.1 Laboratory animal veterinarian shall be in charge of all animals housed and experimented and shall be responsible for ensuring their health and well-being, performing and documenting welfare assessments of all animals in the colony daily.

10.11.2 All animal cages must be checked once a day for signs of illness and/or injury, including weekends and holidays. If animals are being used in experiments in which a sudden decline in health is anticipated, monitoring should be more frequent to ensure the avoidance of unnecessary suffering.

10.11.3 Any signs such as unable to eat or drink, diarrhea, lack of feces or any change in consistency or appearance, bloody urine, etc, or signs of injury must be reported to the veterinarian immediately.

10.11.4 The facility's veterinarian must take a call for treatment or euthanasia for animals.

10.11.5 When there is a sudden death and probable disease, a post-mortem examination should be performed and further diagnostic tests if the veterinarian deems it necessary and shall be documented.

10.11.6 The veterinarian may recommend histopathological evaluation and provide advice as needed to isolate the animal.

10.11.7 The entire colony shall be quarantined if tests are positive for infectious disease.

10.11.8 When a diseased colony (or a suspected disease) is removed, the animal room is disinfected/fumigated before the introduction of another batch of animals.

10.11.9 Organization should have its health monitoring program based on the disease prevalence in that area.

10.11.10 An external contract laboratory or in-house laboratory should be used to perform a health monitoring program of various microorganisms.

10.11.11 The health monitoring reports shall be available in the facility from time to time using sentinel animals.

11 OFFICER-IN-CHARGE OF LABORATORY ANIMAL FACILITY

The person actually in charge of the laboratory

animal facility should be a veterinarian.

12 RECORDS

Suitable records shall be maintained as mentioned below:

- a) Animal breeding and mating;
- b) Stock/census;
- c) Procurement, supply/sale;
- d) Culling;
- e) Feeding;
- f) Environmental variables (*Min/Max* temperature, RH, etc);
- g) Mortality and necropsy records;
- h) Health records of animals and staff;
- j) Laboratory animal facility plans including floor plan and all fixtures, etc; and
- k) CCTV footage register.

13 POTENTIALLY PAINFUL AND INVASIVE PROCEDURES AND HUMANE END POINTS

13.1 Potentially Painful and Invasive Procedures

Invasive procedures shall only be done under strict supervision and guidance of a veterinarian, under general anesthesia and analgesia shall be offered in such cases.

13.2 Humane Endpoints

13.2.1 It shall be the responsibility of each investigator to ensure the avoidance of unnecessary suffering of laboratory animals during experimentation.

13.2.2 To ensure this, the determination and implementation of humane endpoints such as the ones based on weight loss, body temperature, activity, biochemical parameters, corticosteroid levels, etc shall be considered.

13.2.3 Death as an endpoint to a procedure should be avoided as far as possible.

13.2.4 It shall be the veterinarian's responsibility to observe and implement actions to alleviate the pain and suffering of experimental animals based on these humane endpoints.

13.2.5 Where ever the investigator and the veterinarian cannot reach a consensus to act based on humane endpoints to alleviate pain and suffering, the decision of veterinarian shall prevail and be the final.

ANNEX A

(Foreword)

COMMITTEE COMPOSITION

Animal Husbandry and Equipment Sectional Committee, FAD 32

<i>Organization</i>	<i>Representative(s)</i>
Sher-e-Kashmir University of Agricultural Sciences & Technology of Jammu, Jammu	DR BHUPENDRA NATH TRIPATHI (<i>Chairperson</i>)
All India Poultry Breeders Association, New Delhi	DR A. K. RAJPUT DR R. K. JAISWAL (<i>Alternate</i>)
Animal Welfare Board of India, Faridabad	MS PRACHI JAIN DR DEBALINA MITRA (<i>Alternate</i>)
Bihar Animal Sciences University, Patna	DR DEEP NARAYAN SINGH DR RANJANA SINHA (<i>Alternate</i>)
Dau Shri Vasudev Chandrakar Kamdhenu Vishwavidyalaya, Anjora	DR DHIRENDRA BHOSLE DR O. P. DINANI (<i>Alternate</i>)
Department of Animal Husbandry and Dairying, Panchkula	DR BIRENDER SINGH LAURA DR DHARMVIR (<i>Alternate</i>)
Federation of Indian Animal Protection Organizations, New Delhi	DR SIRJANA NIJJAR DR DINESH MOHITE (<i>Alternate</i>)
Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana	DR NAVDEEP SINGH DR SIKH TEJINDER SINGH (<i>Alternate</i>)
ICAR - Central Avian Research Centre, Bareilly	DR JAGBIR SINGH TYAGI DR JAIDEEP ROKADE (<i>Alternate</i>)
ICAR- Central Institute for Research on Buffaloes, Hisar	DR R. K. SHARMA DR SUSHIL KUMAR PHULIA (<i>Alternate</i>)
ICAR - Central Sheep and Wool Research Centre, Avikanagar	DR RANDHIR SINGH BHATT DR SROBANA SARKAR (<i>Alternate</i>)
ICAR - Directorate of Poultry Research, Hyderabad	DR SANTOSH HAUNSHI DR M. NIRANJAN (<i>Alternate</i>)
ICAR - Indian Veterinary Research Institute, Bareilly	DR SUBRATA KUMAR GHOSH DR AMIT KUMAR (<i>Alternate</i>)
ICAR - National Research Centre on Equines, Hisar	DR S. C. MEHTA DR THIRUMALA RAO TALLURI (<i>Alternate</i>)
ICAR - National Research Centre on Pig, Guwahati	DR R. THOMAS DR SUNIL KUMAR (<i>Alternate</i>)
Indian Poultry Equipment Manufacturers Association, Hyderabad	SHRI HARISH RAJARAM GARWARE SHRI ANIL SOMNATH DHUMAL (<i>Alternate</i>)
National Dairy Development Board, Anand	DR R. O. GUPTA DR A. V. HARIKUMAR (<i>Alternate</i>)
National Dairy Research Institute, Karnal	DR ARUN KUMAR MISRA DR SURENDER SINGH LATHWAL (<i>Alternate</i>)
National Egg Coordination Committee, New Delhi	SHRI AJIT SINGHD SHRI BHAGWATI SINGH (<i>Alternate</i>)
National Institute of Animal Nutrition and Physiology, Bengaluru	DR RAVI KIRAN G. DR RAMACHANDRAN (<i>Alternate</i>)

<i>Organization</i>	<i>Representative(s)</i>
PETA India, Mumbai	DR KIRAN AHUJA MS FARHAT U. I. AIN (<i>Alternate</i>)
People for Animals, New Delhi	MS GAURI MAULEKHI MS SHREYA PAROPKARI (<i>Alternate</i>)
Poultry Federation of India, Sonipat	SHRI RANPAL DHANDA SHRI RAHUL KHATRI (<i>Alternate</i>)
Tamil Nadu Veterinary and Animal Sciences University, Chennai	DR S. MEENAKSHI SUNDARAM DR M. R. SRINIVASAN (<i>Alternate</i>)
Uttar Pradesh Pandit Deen Dayal Upadhyaya Pashu Chikitsa Vigyan Vishwavidyalaya Evam Go-Anusandhan Sansthan University (DUVASU), Mathura	DR YAJUVENDRA SINGH DR MUNEENDRA KUMAR (<i>Alternate</i>)
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Member Secretary
SHRI PRADEEP SHARMA
SCIENTIST 'B'/ASSISTANT DIRECTOR
(FOOD AND AGRICULTURE), BIS

Panel on Expert Panel for Review of Standards on Laboratory Animals and Its Quality Management Panel,
FAD 32 : P5

<i>Organization</i>	<i>Representative(s)</i>
ICAR- Indian Veterinary Research Institute, Bareilly	DR PUSHPENDRA KUMAR (<i>Convener</i>)
All India Institute of Medical Sciences, New Delhi	DR P. K. YADAV
Committee for the Purpose of Control and Supervision of Experiments on Animals, New Delhi	DR S. G. RAMACHANDRA
Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana	DR NAVDEEP SINGH
ICMR - National Institute of Virology, Pune	DR DILIP R. PATIL
National Institute of Animal Nutrition and Physiology, Bengaluru	DR N. RAMACHANDRAN
National Institute of Immunology, New Delhi	DR P. NAGARAJAN
PETA India, Mumbai	DR ANKITA PANDEY

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