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***Indian Standard***

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**GOOD AQUACULTURE PRACTICES FOR SEA CAGE**

**FARMING — REQUIREMENTS**

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मानक भवन, 9 बहादुर शाह ज़फर मार्ग, नई दिल्ली - 110002

MANAK BHAVAN, 9 BAHADUR SHAH ZAFAR MARG

NEW DELHI - 110002

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FOREWORD

This Indian Standard was adopted by the Bureau of Indian Standards, after the draft finalized by the Fish, Fisheries and Aquaculture Sectional Committee had been approved by the Food and Agriculture Divisional Council.

In recent times, marine fish catch in the country is stagnating annually, indicating that increasing the fish catch from the existing fishing grounds is not sustainable economically and ecologically. Added to this, dwindling catch in capture fisheries, resulting in unemployment in the coastal region and demand for additional seafood necessitates the development of mariculture as a substantial seafood production sector. Enhancing fish production from the inland sector has limited scope and the major portion of the additional demand has to come from mariculture. Sea cage farming is viewed as a major option for increasing seafood production and has been expanding rapidly in recent years at the global level. The cage aquaculture has grown very rapidly during the past 20 years and is presently undergoing rapid changes due to globalization and growing global demand for aquatic products.

In this context, it was felt necessary to bring out standardized guidelines for good aquaculture practices in sea cage culture for the fishermen, fish farmers and officials of the fisheries departments. Through this guide, the concerns about food safety and quality, sustainability, environmental protection, etc. are being catered to. It is necessary to pay attention to the quality of production practices requiring minute attention to different aspects of production, storage, handling and distribution. The cage-farmed fish include mostly cobia, pompano, grouper, Asian seabass and snappers in our country.

This Standard is part of a series of Indian Standards on Good Aquaculture Practices. Considerable assistance has been provided by ICAR-Central Marine Fisheries Research Institute, Kochi, in the formulation of this standard.

The composition of the Committee and the panel responsible for the formulation of this standard is given in Annex A.

For the purpose of deciding whether a particular requirement of this standard is complied with, the final value, observed or calculated, expressing the result of a test or analysis, shall be rounded off in accordance with IS 2 : 2022 ‘Rules for rounding off numerical values (*second revision*)’. The number of significant places retained in the rounded off value should be the same as that of the specified value in this standard.

***Indian Standard***

**GOOD AQUACULTURE PRACTICES FOR SEA CAGE FARMING — REQUIREMENTS**

**1 SCOPE**

This Indian Standard provides Good Aquaculture Practices for cage culture in seawater and associated products and procedures like cages, site and species selection, seed, feed and health management, safety measures, market and post-harvest facilities.

**2 REFERENCES**

The following standards contain provisions which through reference in this text, constitute the provision of this standard. At the time of publication, the editions indicated were valid. All standards are subject to revision and parties to agreements based on this standard are encouraged to investigate the possibility of applying the most recent editions of the standards indicated below:

|  |  |
| --- | --- |
| *IS No.* | *Title* |
| 16150 (Part 6) : 2023 | Fish feed — Specification, Part 6 Marine Carnivorous fish feed |

**3 GOOD AQUACULTURE PRACTICES FOR CAGE CULTURE IN SEAWATER**

**3.1** **Site Selection**

Appropriate site selection decides the profitability and sustainability of the cage farm. The sites selected, directly impact construction costs, operating costs, growth and survival rate of fish, and shelf-life of cage accessories. Before the establishment of cage farming, an extensive knowledge of the site environment (topography and water quality parameters) is required by involving detailed surveys, secondary information from existing literature and government sources and first-hand information from local people.

Major criteria to be followed for site selection are topographical, physical, chemical, biological, accessibility, social problem and legal aspects.

**3.1.1** *Topographical Parameters*

1. Cages shall be installed in sheltered areas, protected from strong winds and waves; in general, wind velocity and height of wave should preferably not exceed 10 knots and 1.0 m for floating cages.
2. The preferred depth for a floating cage is 8-10 m. Sufficient depth beneath the cage is necessary to avoid oxygen depletion, and accumulation of uneaten food, faeces and debris under the cage.
3. A firm substrate, with a combination of fine gravel, sand and clay is an ideal site for cage culture.

**3.1.2** *Physical Criteria*

1. Tidal currents help to keep the cage site clean, bring fresh oxygenated water and remove waste from the cage site. The ideal current speed should be 50-100 cm/sec, and should not exceed 100 cm/sec.
2. Turbid water affects the visibility of the feed to fishes, and if turbidity remains for a longer duration, then it results in reduced feed intake and impaired fish growth.
3. Suitable temperature for the optimum metabolic activities of fish ranges from 27-31°C.

**3.1.3** *Chemical Criteria*

Recommended water quality criteria for sea cage culture of marine finfishes are given in Table 1.

**Table 1 Water Quality Criteria for Sea Cage Culture of Marine Finfishes**

(*Clause* 3.1.3)

|  |  |  |
| --- | --- | --- |
| **Sl. No.** | **Water Quality Parameters** | **Optimum Range/ Level** |
| (1) | (2) | (3) |
|  | Dissolved Oxygen | 5– 8 mg/l |
|  | Water Temperature | 27 – 31°C |
|  | *p*H | 7.5 – 8.5 |
|  | Salinity | 15 – 35 % |
|  | Transparency, *min* | 40 cm |
|  | Ammonia, *max* | 0.05 ppm |
|  | Nitrate, *max* | 200 mg/l |
|  | Nitrite, *max* | 4 mg/l |
|  | COD, *max* | 3 mg/l |

**3.1.4** *Biological Criteria*

The presence of more biological organisms will directly affect the shelf-life of the cage accessories. The attachment of these organisms to the cage net or other cage accessories is dependent on the silt accumulation. The silt accumulated in the cage net acts as the substrate for fouling organisms. Therefore, the site selected for cage culture should be free from silt accumulations.

**3.1.5** *Accessibility*

Cage culture activity involves frequent movement between the cage site and shore for feeding, transportation of seed and cage accessories, and routine cage monitoring. Therefore, the cage site should have jetty facilities for ease of voyage. Selected cage sites should have road facilities for the transportation of seed, feed and harvested fish.

**3.1.6** *Social factors*

Social issues are non-biological and one of the major constraints in cage culture activity. Therefore, selected sites should be away from fishing villages, and cage culture areas should be clearly demarcated.

**3.2 Types of cage**

Floating-type cage is commonly used in marine waters. It is supported by a floating frame, where net bags are kept hanging in water without touching the basin. It is generally practiced in water bodies with a depth of water more than 6 m.

**3.3 Cage Design - Size and Shape**

The cage design for fish culture should be able to withstand rough sea conditions, should provide a conducive condition for the fish being reared and should have good maneuverability. For farming along the coasts of India, the circular shape of cages is suggested. Cage size determines operational expenditures and profit. The cages of 6.0 m in diameter and circular in shape are being used with ease in maintenance and providing moderate income, however, larger-sized cages can be used for higher production levels.

The materials used in sea cage farming should be strong enough to withstand rough sea conditions and should be also rust-proof with long durability in seawater. A cage frame made using High Density Poly Ethylene (HDPE) (PE 100) is considered the most suitable for marine cage farming.

**3.4 Cage Accessories**

Different cage accessories are being used for complete cage fabrication and deployment. The different accessories are a cage frame, mooring system, ballast system and cage net.

**3.4.1** *HDPE Cage Frame*

The cage frame has two collar rings with floatation properties and a middle catwalk, and all three frames assist in routine cage management.

1. Inner and outer collar rings, with 6.0 m and 7.0 m diameter respectively, are made up of HDPE pipes of 140 mm pipe outer diameter. The middle catwalk, with a 6.5 m diameter, is made up of HDPE pipe with a 90 mm pipe outer diameter. The inner and outer collar pipes are filled with polyurethane foam or thermocol to enhance their floatation efficiency.
2. Base pipes help to keep inner and outer collar rings together and provide shape for the cage. Base pipes are connected to the handrail via vertical and diagonal supports. Base pipes are made up of HDPE pipes with a 250 mm pipe outer diameter.
3. Vertical and diagonal supports help to join the base collar rings to the handrail. They are made up of HDPE pipes with a 90 mm pipe outer diameter.
4. The handrail is used for tying the inner net, and during routine cage management, it functions as a support for the workers. The handrail is made up of HDPE pipes with a 90 mm pipe outer diameter.

**3.4.2** *Cage Mooring System*

1. The mooring system holds the cage in the desired position and depth with mooring chains, and anchors. Individual cages are to be moored using single-point mooring and a battery of cages should be moored through grid mooring.
2. The use of anchors in the mooring system depends on the sea bottom. The preferred anchors along the east-coast are concrete blocks or stones. For sites with clayey bottom, toothed anchors along with concrete blocks are suggested.
3. Use of concrete cement blocks is the preferred anchoring system. Concrete blocks, each weighing approximately 200 kg are used, and 10-12 such blocks, with a total weight of 2.0 to 2.5 tonnes are recommended for effective and efficient anchoring.
4. Mooring chain connects the anchoring system to the cage frame via floats (buoy). Long-linked alloy steel chain of 14.0 mm outer diameter and 22 tonnes shearing strength is preferred. If the chain thickness is less than 14.0 mm, it gets easily eroded and may not hold the tensions associated with rough sea. Two such mooring chains are used for each cage and are secured with the help of D-Shackles. For a site with a 10.0 m water depth, approximately 100 m of mooring chain is required for ease of cage movement.
5. In addition to providing floatation, buoys act as shock absorbers in the mooring system. The pressure created on the mooring chain by currents and winds is prevented by the buoys from directly impacting the cage frame. Three buoys of 200 litre capacities are used. Buoys are coated with Fiber Reinforced Plastic (FRP) to prevent rusting of clamps.
6. D-Shackles help in securing the mooring chain to the concrete blocks, swivel, buoy and cage frame. D-Shackles, made of stainless steel with 19.0 mm outer diameter, are used to counteract the heavy load. A mooring system with 10-12 numbers of 200 kg cement blocks and 100 meter of mooring chain requires nearly 35-40 D-Shackles.
7. Swivel is attached to the middle of the mooring chain and helps to rotate the cage frame freely in different directions in tune with the water current.

**3.4.3** *Ballast System*

1. Ballast pipes help to maintain the cage net structure intact, and in proper shape against the water movement.
2. Ballast is prepared using a 2.5 cm diameter perforated HDPE pipe. To ensure maximum net space to allow sufficient fish movement, the inner net is tied with two ballast pipes, at the bottom and middle, and this prevents the net from getting distorted due to constant water movement.
3. For sites with turbulent sea conditions, steel or iron rods are inserted within perforated HDPE pipes to provide strength to the ballast. However, usage of steel or iron rods should be avoided as far as possible, because if rods come out of HEPE pipes, they may tear the net.
4. While mooring cages, the ballast pipes should be shifted along with the cages to avoid extra expenditure on separate shifting.

**3.4.4** *Cage Net*

1. HDPE braided nets are preferred for their strength and light weight. HDPE nets are durable in seawater for up-to 5 years. In cages, three nets are used: outer net, inner net and bird net. The inner and outer net of sapphire blue is the preferred colour.
2. Outer net is vertically hung from the outer collar. It functions to prevent the entry of predators to the inner net. Net with a mesh size of 40.0 mm should be used, both for avoiding predator entry and for providing a relatively lesser load on the cage frame. Optimum size of the outer net is 7.0 m in diameter and 4.0 m in depth. Mesh sizes less than 40.0 mm burden the cage frame with additional load, and thus should be avoided. For preparing the specified net size, a raw net weight of 18-20 kg is required.
3. Inner net is vertically hung from the handrails of the cage frame. It functions to hold the cultured fish within the cage net structure. Inner net of 25.0 mm mesh size is used for rearing fishes above 100g in size. However, mesh size is also dependent on the shape of the fish. Indian pompano is broad in shape, whereas orange-spotted grouper is cylindrical; therefore, the chances of escape for Indian pompano are much less when compared to orange-spotted grouper for any specific mesh size. Despite the above, for fish of 100 g size, an inner net with 25.0 mm mesh size is ideal. Fishes whose width is less should initially be stocked in smaller mesh (1.5 or 2.0 mm) net until they reach the optimum size. The use of a 25.0 mm mesh net is recommended for the free flow of water and to reduce fouling accumulation. Fouling accumulation is more in small mesh net due to the availability of more surface area for attachment. The optimum size of the inner net is 6.0 m in diameter and 5.0 m depth. For preparing the specified net size, a raw net weight of 25-30 kg is required.
4. Bird net is tied to the handrails and is placed horizontally. It helps to prevent fish predation by birds. Nylon net of 1.25 mm twine thickness and 80 mm mesh size is preferred for preventing birds’ predation.
5. Specification for inner net mesh size in different grow-out stages for marine finfish is given in Table 2.

**Table 2 Specification for Inner Nets Mesh Size in Different Grow-Out Stages for Marine Finfishes**

[*Clause* 3.4.4 (e)]

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Sl No.** | **Species** | **18 mm Mesh [Fish Size (g)]** | **25 mm Mesh [Fish Size (g)]** | **40 mm Mesh [Fish Size (g)]** | **60 mm Mesh [Fish Size (g)]** |
| (1) | (2) | (3) | (4) | (5) | (6) |
|  | Cobia | 10-70 | 100-1100 | 1100-4000 | 4000-7000 |
|  | Pompano | 10-100 | 100-1000 | 1000-2000 | - |
|  | Grouper | 15-300 | 300-1000 | 1000-2000 | - |
|  | Seabass | 15-300 | 300-1000 | 1000-2000 | - |

**4 CAGE STRUCTURE DEPLOYMENT**

Deployment of mooring systems and cage frame structure should be done one after the other with the help of boats. The mooring system should be deployed at least one week before the deployment of the cage frame, and this time gap is required for the proper setting of the anchoring system in the sea bottom.

**4.1 Mooring Systems Deployment**

1. Concrete blocks of required numbers are arranged at distances of 1.0 m apart and are connected via a mooring chain with the help of D-shackles. After blocks, the chain is connected to the swivel and finally to the three buoys. The buoys are placed at distances of 2.0-3.0 m apart.
2. The entire structure is loaded onto the vessel (trawlers) and arranged in an order at the rear end. At the selected location, the entire structure is dropped into the water, and due to the anchor weight, the mooring system settles in the location.
3. The attached buoys help for floatation and also for identifying the mooring locations.

**4.2 Deployment of Cage Frame**

1. The cage frame is deployed with the help of outboard motor vessels. The frame is tied to the boat with the help of 8.0-10.0 mm rope and is dragged to the selected site, where it is attached to the mooring chain after the buoys. The distance between the first buoy and the cage frame should be at least 3.0 m.
2. The ballast pipes are also shifted with the cage frame by attaching to the handrails.

**5 SPECIES SELECTION**

1. Selection of fish species is pivotal for cage culture operation. While selecting the species for culture, certain criteria like biological, economic aspects, and consumer preference should be given prime importance for economic sustainability.
2. The major criteria to be considered before selecting the species are sufficient availability of fish seeds, availability of hatchery technology for seed production in a confined environment, acceptance to artificial feeds, tolerance to different environmental conditions, compatibility to culture in various systems, resistance to disease and stress, high nutritional value, consumer acceptance, economic value in local and international market and regional preference.
3. As per the above criteria, Indian pompano (*Trachinotus mookalee*), Cobia (*Rachycentron canadum*), Silver Pompano (*Trachinotus blochii*), Groupers (*Epinephelus sp*.), Seabass (*Lates calcarifer*), and Snappers (*Lutjanus* sp.) are highly suitable for sea cage farming.

**5.1 Fish Seed**

The quality of fish seed is of vital importance for the success of grow-out culture in cages. Uniform-sized seeds appropriate for the mesh size of the fish net cage should be stocked to prevent their escape. This will also help in selecting the correct sized feed for fishes, avoiding wastage of feed and reducing cannibalism. Seeds should be healthy, and free from diseases and deformities. Fish seed specifications for different species of marine finfish are given in Table 3.

**Table 3 Fish Seed Specifications for Different Species of Marine Finfishes**

(*Clause* 5.1)

|  |  |  |
| --- | --- | --- |
| **Sl. No.** | **Aspects** | **Specification** |
| (1) | (2) | (3) |
|  | Stocking Size | Marine Fin fishes: ≥ 20 – 25 g (~10-15 cm) |
|  | Seed Quality | 1. Large effective population size of broodstock fishes (Ne): At least 50 broodfish pairs. 2. Permissible level of inbreeding accumulation: ~1% per generation. 3. Family Selection based breeding. Use of Factorial Mating design. 4. Fish seed certification system preferred. |
|  | Seed Purity | 1. Monoculture of preferred/desirable fishes at hatchery level 2. Community breeding of multiple fish species in the same breeding tank must be stopped. 3. Fish seed certification system preferred. |
|  | Stocking Strategy | 1. Should be transported in an open container with dissolved oxygen support and oxygen concentration should be above 5.0 ppm 2. Stocking uniform size is preferred to avoid cannibalism during grow-out culture operation. |

**6 NURSERY REARING**

**6.1** Rearing fish larvae through the early life stages is performed in the nursery, and this is the phase between hatchery and grow-out. Thus, before stocking for grow-out, culture species need to be nursed for attaining optimum stocking size.

**6.2** Nursery rearing of marine fin fishes is essential in cage culture for reducing the culture duration during grow-out. Three types of nursery systems are preferably used: hapa-based nursery in earthen ponds, recirculating aquaculture system (RAS) based nursery and concrete or FRP tank-based flow-through nursery. These nursery facilities should be established near to cage site for ease of fish transfer.

**6.3** Feed used in the nursery should be appropriate for fish species. Feeding frequency of 4-5 time/day at 8-12% body weight is recommended. The feeding rate varies with the size of the fingerlings and species reared.

**6.4** For the majority of the species, with proper nursery management, advanced fry of 2.5 cm (0.5 to 0.6 g) stocked at 500 nos/m3 should reach 6.25 cm (5.0 g) within 45 days. Optimum feed size to avoid size variation should be 0.8 to 1.2 mm, at 12% of body weight. During the later phase of nursery, early fingerlings stocked at 5.0 g size should attain 25.0 g size in 30 days at a stocking density of 300/m3 with a feeding rate of 10% body weight. Therefore, during the entire nursery duration, advanced fry of 2.5 cm size should reach 25.0 g in two and half months (75 days).

**6.5** Indian pompano, being a fast-moving pelagic fish, dissolved oxygen requirement is very high; therefore, during nursery, the dissolved oxygen concentration should always be above 4.5 ppm.

**6.6** With proper feeding and water quality management, expected survival in RAS and indoor tank-based cultures should be above 96%, whereas in hapa-based earthen ponds, more than 90-95% survival is expected.

**6.7** Fishes are very active during nursery rearing; therefore, they tend to jump to at least 15.0 cm above the water level. Thus, the water level should be at least 30.0 cm below the tank surface to avoid fish fingerlings falling out of water. It is suggested to cover the tank surface with fish net to avoid fish jumping out of the tank.

**6.8** Vibriosis is the most common bacterial infection occurring during nursery, because of stress. Minimizing stress in the nursery will help to keep the fishes free from bacterial infection. Possible stressors are: overcrowding, more waste accumulation in tank bottom, rough handling, higher water temperature and lower dissolved oxygen.

**7 SEED TRANSPORTATION FOR GROW-OUT STOCKING**

**7.1** It is preferred to establish the nursery unit near to cage culture site for ease of transportation. Advanced fingerlings reared in the nursery should be transferred to the cage site either in polythene bags filled with oxygen or in plastic or FRP tanks supported with oxygen.

**7.2** When fingerlings are stocked at more than 5g in size, they should be transported in a container supported with pure oxygen to achieve maximum survival. Tanks or open container-based seed transportation with oxygen supply is the preferred mode of transportation for better survival.

**7.3** Adequate care should be provided while transferring fingerlings. Fingerlings transported in stressed condition (overcrowding and less dissolved oxygen) are more susceptible to vibriosis after stocking in cages. Thus, adequate care should be given to keep the animals under stress-free conditions.

**7.4** Use of ice while seed transportation is recommended during the summer season to avoid heat shock to the transported larvae.

**7.5** Seeds transported over long distances should be in sintex tanks supported with oxygen, and for short distances of less than an hour, in open FRP tanks supported with oxygen.

**7.6** Based on the current research; the optimum fish size, stocking density and mode of transportation are given in Table 4.

**Table 4 Transportation of Marine Finfish Seeds**

(*Clause* 7.6)

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Sl. No.** | **Fish Size (g)** | **Duration**  **(hr)** | **Stocking**  **(nos/l)** | **Mode of transportation** |
| (1) | (2) | (3) | (4) | (5) |
|  | > 0.25 | 24-36 | 50-60 | Polythene bag filled with oxygen |
|  | 1.0 to 2.5 | 15-30 | 20-25 | Polythene bag filled with oxygen |
|  | 2.5 to 5.0 | 12-24 | 10-15 | Sintex tank supported with oxygen |
|  | 5.0 to 15.0 | 12-20 | 4-5 | Sintex tank supported with oxygen |
|  | 25.0 to 30.0 | 12-20 | 1-1.5 | Sintex tank supported with oxygen |

**8 STOCKING**

Stocking the appropriate size and number of fish seed in cages is very crucial for the success of cage farming. After allowing the hatchery-produced spawn to grow for a period ranging from 30 to 60 days, fish seed can be stocked in cages. Nursery rearing of seed is essential for all species and it can be done as a separate activity, in land-based nursery ponds or hapas held in ponds or in floating nursery cages, by individuals or groups at different localities to support sea cage farming with ready-to stock fingerlings. Healthy, uniform-sized fingerlings should be procured for stocking in cages. The fingerling stocking details are given in Table 5.

**Table 5 Optimum Stocking Size and Stocking Density for Different Species of Marine Finfishes**

(*Clause* 8)

|  |  |  |  |
| --- | --- | --- | --- |
| **Sl No.** | **Species** | **Stocking Size**  **(Length/Weight)** | **Stocking Density (No/m3)** |
| (1) | (2) | (3) | (4) |
|  | Cobia | 15 cm/35 g | 8-10 |
|  | Silver Pompano | 10 cm/35 g | 30-40 |
|  | Indian Pompano | 11 cm/35 g | 25-30 |
|  | Orange Spotted grouper | 12 cm/35 g | 20-25 |
|  | Asian Seabass | 15 cm/35 g | 30-40 |

**9 GROW-OUT CULTURE**

The requirements for ensuring the quality of grow-out culture are as follows:

1. After reaching the cage site, transported juveniles should be slowly released into the cage for acclimatizing to the cage water environment.
2. Appropriate stocking density should be followed to achieve optimum growth and economic benefit.
3. Artificial floating pelleted feed with high nutritional composition is recommended for grow-out systems.
4. Feed should be broadcasted in the middle of the cage. While broadcasting the feed, some feed gets wasted by drifting due to wave and wind action. Therefore, attaching a feed mesh, of 0.10-0.05 mm mesh size and 1.0 m width, to the inner net helps in controlling the feed wastage. Even, using mosquito net can act as feed mesh for controlling feed wastage.
5. It is suggested to broadcast feed slowly and at multiple times during each feeding. This will ensure equal availability of feed for all fish, and fish deprived of feed will be non-existent.
6. Recommended feeding rate in grow-out culture varies from 6.0-1.5%, according to the growth of the fish, and this ration should be divided and given 3.0-4.0 times during each day.
7. Better feed digestion and assimilation ensure better fish growth, thus, a minimum of 3 hours’ time interval between two feeding schedules should be given, and therefore, the feeding frequency should be decided accordingly. However, feeding should be provided at least twice a day to maintain good fish health. Feeding frequency of 3-4 times/day has been observed to show better growth instead of feeding twice.
8. In grow-out culture, fish growth should be monitored fortnightly and the feeding rate to be adjusted based on the weight gain after every sampling.
9. In a well-managed cage culture grow-out system, fish fingerlings stocked at 20 to 25g require nearly 10 months to reach the market size of 800-1000 g, whereas if it is stocked at 100 g size, it takes 5 to 6 months to reach the same size.
10. Survival rate varies with stocking size; survival rate for fish stocked at 25 g varies from 90 to 95 %, whereas for fish stocked at 100g, survival ranges between 95 to 98 %.
11. Feed conversion Ratio (FCR) varies from 1.65 to 2.0, and achieving low FCR is dependent on effective feed management.
12. Fish growth, feeding frequency and rate of feeding for Indian pompano in marine cage culture are given in subsequent clauses.

**10 FEEDING**

Any material used for feeding should contain the following five principal constituents; Protein, Carbohydrate, Fat, Minerals and Vitamins, in appropriate balance. For feeding of marine carnivorous fish, fish feed conforming to IS 16150 (Part 6) is recommended.

**10.1 Feeding Storage**

During storage, feeds undergo deteriorative changes which not only lower their nutritive value but also affect their palatability and appearance. Feeds should be stored in dry ventilated warehouses away from direct sunlight at more or less constant temperatures. All feeds should be used within the prescribed time and inspected regularly. During long storage, there may be changes such as fungal growth, degradation of vitamin potency and fat rancidity. Unnecessary handling may damage feed bags and reduce pellets to powder that is usually not consumed by fish and wasted. Pests like rats, cockroaches, etc., must be strictly controlled in the storage, to avoid contamination. Proper storage of feed is simple, but it is important to keep its safety and quality.

**10.2 Feeding Schedule**

Feeding rates, frequency of feeding and time of feeding are important factors to be considered in cage farming. Feeding rates and frequencies are related to the age and size of the fish. Fish larvae and fry need to be fed on a high-protein diet more frequently. When fishes grow bigger, feeding rates and frequencies can be reduced. Feeding fish is a labour-intensive activity and the frequency has to be adjusted in such a way that it is economically viable. Generally, growth and feed conversion increase with the increase in feeding frequency. Feed consumption is also influenced by time of day, season, water temperature, dissolved oxygen levels and other water quality parameters. Even though several feeding charts are available, it is better to construct one of your own with information on: Days of Culture (DOC), Fish Weight, Protein in Feed, Meal/Day, Feed Consumed as % of body weight, Average Daily Gain (ADG) and Feed Conversion Ratio (FCR). Indicative feeding charts for Indian pompano, orange-spotted grouper, Silver Pompano and Cobia are given in Table 6, Table 7, Table 8 and Table 9 respectively.

**Table 6 Growth, Feed and Feeding for Indian Pompano**

(*Clause* 10.2)

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Sl. No.** | **DOC** | **Size (g)** | **Feed Size (mm)** | **Feeding**  **Rate (%)** | **Feeding Frequency**  **(times/day)** |
| (1) | (2) | (3) | (4) | (5) | (6) |
|  | 0-30 | 25-50 | 1.2 to 1.8 | 8 | 4-5 |
|  | 30-120 | 50-100 | 1.8 to 3.0 | 6-5 | 4-5 |
|  | 120-180 | 100-300 | 3.0 to 4.0 | 5-4 | 4 |
|  | 180-210 | 300-500 | 4.0 to 6.0 | 4-3 | 4 |
|  | 210-300 | 500-750 | 6.0 to 7.0 | 3-2.5 | 4-3 |
|  | 300-360 | 750-1100 | 7 to 10.0 | 2 | 3 |

**Table 7 Growth, Feed and Feeding for Orange Spotted Grouper**

(*Clause* 10.2)

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Sl. No.** | **DOC** | **Size (g)** | **Feed Size (mm)** | **Feeding Rate (%)** | | **Feeding Frequency (times/day)** |
|  |  |  |  | Artificial  feed (A.F) | A.F + Trash fish |  |
| (1) | (2) | (3) | (4) | (5) | (6) | (7) |
|  | 0-60 | 20-75 | 1.8 to 3.0 | 8% | 4% + 5% | 4 |
|  | 60-120 | 75-150 | 3.0 to 5.0 | 6-5% | 3% + 5% | 4 |
|  | 120-180 | 150-275 | 5.0 to 6.0 | 5-4% | 2% + 5% | 2 |
|  | 180-240 | 275-450 | 6.0 to 1.0 | 4% | 2% + 5% | 2 |
|  | 240-300 | 450-650 | 1.0 to 1.5 | 3-2.5% | 1% + 5% | 2 |
|  | 300-360 | 650-900 | 1.0 to 1.5 | 2% | 1% + 5% | 2 |

**Table 8 Growth, Feed and Feeding for Silver Pompano**

(*Clause* 10.2)

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Sl. No.** | **DOC** | **Size (g)** | **Feed size (mm)** | **Feeding rate (%)** | **Feeding frequency (times/day)** |
| (1) | (2) | (3) | (4) | (5) | (6) |
|  | 0 to 30 | 15 to 35 | 1.2 to 1.8 | 8 | 4 to 5 |
|  | 30 to120 | 35 to 172 | 1.8 to 3.0 | 6 to 5 | 4 to 5 |
|  | 120 to180 | 172 to 375 | 1.8 to 3.0 | 5 to 4 | 4 |
|  | 180 to 210 | 375 to 465 | 3.0 to 4.0 | 4 to 3 | 4 |

**Table 9 Growth, Feed and Feeding for Cobia**

(*Clause* 10.2)

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Sl. No.** | **DOC** | **Size (g)** | **Feed Size (mm)** | **Feeding Rate (%)** | **Feeding Frequency**  **(times/day)** |
| (1) | (2) | (3) | (4) | (5) | (6) |
|  | 0-30 | 35-70 | 1.5 to 1.8 | 15 | 4-5 |
|  | 30-120 | 70-460 | 1.8 to 2.5 | 15-10 | 4-5 |
|  | 120-180 | 460-1900 | 2.5 to 5.0 | 10-8.0 | 4 |
|  | 180-210 | 1900-3300 | 5.0 to 8.0 | 8.0-5.0 | 4 |
|  | 210-300 | 3300-5600 | 12.0 to 15.0 | 8.0-5.0 | 4-3 |
|  | 300-360 | 5600-7200 | 15 to 18.0 | 5.0-3.0 | 3 |

**11 CAGE STRUCTURE MANAGEMENT**

**11.1** Managing cage frame and other accessories is an important component of marine cage culture. Periodical monitoring of cage frame and accessories helps in avoiding loss of fish stocks due to escape caused by net damage, and because of cage drifting away from mooring. Proper and routine monitoring will also help improving the shelf life of the cage.

**11.2** Fish fingerlings stocked at 25g size require a culture duration of ten months; therefore, different cage components should be managed efficiently. Various management activities include net exchange, cage frame cleaning and mooring checking.

**11.3** The cage net is the structure that holds the fish and is prone to attachment of barnacles, and algal and silt accumulation. Thus the net needs to be exchanged periodically, depending on the accumulation rate. The accumulation depends on the season and the location, and based on the experiences from the east coast, the cage net should be exchanged at least once in two months. If the cage nets are not exchanged within the stipulated time, then, they may tear off due to the heavy load. Also, fouling load on the cage net will negatively impact the buoyancy of the cage frame.

**11.4** Cage frame, being the walkway, is prone to settlement of barnacles, and if the settlement is more, it would adversely impact the shelf life of the cage. Also, settlement in cage frame, more often than not results in the tearing of the net ropes through rubbing. The attachment of micro and macroalgae (*Ulva* spp) in the cage frame leads to skidding during routine management. Thus, the cage frame requires monthly cleaning.

**11.5** Cage mooring helps to keep the entire cage structure in position, thus the mooring chain requires continuous monitoring, at least once a month. The mooring system specified for the cages will remain without much of an issue for a minimum of two years, and then slowly the chain starts eroding, resulting ultimately in chain tear. Thus, mooring checking with the help of an underwater diver is recommended once a month. With prevailing heavy wind and waves along the east coast of India, the chains are mostly damaged in proximity to the anchors, below the swivel; therefore, providing a single additional chain of 10 m length between anchors and swivel will result in an additional time of shelf life for the mooring chain.

**12 FISH HEALTH MANAGEMENT**

Disease outbreaks can occur as a result of intensive farming in cages. Infectious diseases are mainly due to waste accumulation, crowding, handling, variations in water quality parameters and bio-fouling. Cage abrasion can cause fin and skin damage to farmed fish. Occurrence of infections/ diseases can be minimized by selecting a good site, optimal stocking density and careful handling of fish stock. Fish farmers should maintain a record of weather, water quality parameters, feeding rate, length and weight of fish sampled, fish behavioral changes, net cage exchange details, etc. These records provide useful information for analysis of health status. The important aspects that should be followed for disease control in cage culture systems are as follows:

1. Cage-cultured fish should be checked and critically observed for their feeding and health status by periodic sampling at fortnight intervals.
2. Also, daily observations during feeding are essential for understanding the feeding behaviour, which is an excellent indicator of the health status of the fish.
3. The major diseases associated with the grow-out cage culture of pompano are: Vibriosis caused by selected species of *Vibrio* bacteria, and parasitic infestations caused by ecto-parasites. Fish affected by Vibriosis exhibits the symptoms of moving on the water surface, and the eyes and the fins become reddish in colour. Fishes, when infected, do not accept feed and virtually stop feeding. Vibriosis in fish is controlled by the use of prescribed medicated feeds and probiotics.
4. In parasitic infestation, visible minor ulceration appears on the entire body and, importantly in the gills. Isopods cause coin-shaped wounds, mostly in the dorsal side. Also, parasitic attachment can be noticed on the body surface, ultimately killing the fish.
5. The parasitic infestation is controlled by a freshwater dip or by using medicated feeds. Avoiding high stocking density helps to get rid of parasitic infestations in cages.

**13** **FISH HARVEST AND MARKETING**

**13.1** In cages, fishes are reared in small confinement, so harvesting cage-cultured fish is easier than any other culture method.

**13.2** It is suggested to harvest the fish either in the early morning or late evening hours to maintain their freshness.

**13.3** While harvesting, the inner cage net is lifted from all sides, and the lifted net is hung on handrails and tied to it. The fishes in the inner net are harvested with the help of a hand scoop net.

**13.4** Immediately upon harvest, washing in clean water and chill killing is suggested to maintain the freshness and quality of the harvested fish.

**13.5** Harvested fishes should be packed in plastic trays or thermocol boxes, by adding layers of ice in equal quantities below and above the fish.

**13.6** A well-managed cage can harvest an average of 2.0 tonnes of marine fishes /cage/year.

**13.7** Cage-farmed fishes could be harvested based on demand and are most preferably harvested either during the lean fishing period or during the trawl-ban season. The demand during the trawl-ban season is exceptionally high for the species.

**ANNEX A**

(*Foreword*)

**COMMITTEE COMPOSITION**

**FISH, FISHERIES, AND AQUACULTURE SECTIONAL COMMITTEE, FAD 12**

|  |  |  |
| --- | --- | --- |
| *Organization* |  | *Representive(s)* |
| Indian Council of Agricultural Research, New Delhi |  | DR JOYKRUSHNA JENA **(*Chairperson*)** |
| All India Shrimp Hatchery Association, Hyderabad |  | DR JOSHI K. SHANKAR |
|  | SHRI K. MADHUSUDAN REDDY (*Alternate*) |
| Central Agricultural University, Imphal |  | SHRI RATAN KUMAR SAHA |
|  | SHRI ARUN BHAI PATEL (*Alternate I*) |
|  | DR NARESH KUMAR MEHTA (*Alternate II*) |
| Defence Food Research Laboratory, Mysore |  | DIRECTOR |
|  |  | DR RADHIKA. M (*Alternate*) |
| Export Inspection Council of India, New Delhi |  | SHRI N. PALANIKUMAR |
|  |  | SHRI SACHIN PANWAR (*Alternate*) |
| Fisheries College and Research Institute, Thoothukudi |  | DR R. JEYA SHAIKLA |
|  | DR R. SHALINI (*Alternate*) |
| ICAR - Central Inland Fisheries Research Institute, Kolkata |  | DR BIJAY KUMAR BEHERA |
|  | SHRI DHARMENDRA KUMAR MEENA (*Alternate*) |
| ICAR - Central Institute for Fisheries Technology, Kochi |  | DR ZYNUDHEEN A. A. |
|  | DR C. O. MOHAN (*Alternate I*) |
|  | DR T. V. SANKAR (*Alternate II*) |
| ICAR - Central Institute of Brackish Water Aquaculture, Chennai |  | DR K. AMBASANKAR |
|  | DR SUBHENDU KUMAR OTTA (*Alternate I*) |
|  | DR P. K. PATIL (*Alternate II*) |
| ICAR - Central Institute of Fisheries Education, Mumbai |  | DR B. B. NAYAK |
|  | DR SANATH KUMAR (*Alternate*) |
| ICAR - Central Institute of Fresh Water Aquaculture, Bhubaneswar |  | DR KEDAR NATH MOHANTA |
|  | DR P. C. DAS (*Alternate*) |
| ICAR - Central Marine Fisheries Research Institute, Kochi |  | DR A. K. ABDUL NAZAR |
|  | DR SHUBHADEEP GHOSH (*Alternate*) |
| Ministry of Fisheries, Animal Husbandry and Dairying, Department of Animal Husbandry and Dairying, New Delhi |  | DR NILESH ANIL PAWAR |
| National Institute of Fisheries Post Harvest Technology and Training, Ernakulam |  | DR SHINE KUMAR C. S. |
|  | DR REMYA KUMARI K. R. (*Alternate*) |
| IN PERSONAL CAPACITY  [*Old Kalibari Road, Krishnanagar, Agartala, (West) — 799001*] |  | DR R. K. MAJUMDAR |
| BIS Directorate General |  | SHRIMATI SUNEETI TOTEJA, SCIENTIST ‘F’ AND HEAD (FOOD AND AGRICULTURE)  (REPRESENTING DIRECTOR GENERAL (*Ex-Officio*) |

*Member Secretary*

SHRI DEBASISH MAHALIK

Scientist ‘C’/DEPUTY DIRECTOR,

(FOOD AND AGRICULTURE), BIS