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BUREAU OF INDIAN STANDARDS

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भारतीय मानक मसौदा

कार्यस्थल पर वायु — कणिका लेड और लेद के यौगिक ज्ञात करना - ज्वाला अथवा विघुततापीय परमाणु अवशोषण स्पेक्ट्रोस्कोपी विधि

(*IS 15309 का पहला पुनरीक्षण*)

Draft Indian Standard

Workplace Air — Determination of Particulate Lead and Lead Compounds — Flame or Electrothermal Atomic Absorption Spectrometric Method

(First Revision of IS 15309)

(ICS 13.040.30)

Air Quality Sectional Committee, CHD 35	**Last Date for Comments: 30th October 2024**

Air Quality Sectional Committee, CHD 35

NATIONAL FOREWORD

(*Formal clause shall be added later*)

The health of workers in many industries, for example, mining, metal refining, battery manufacture, construction, is at risk through exposure by inhalation of particulate lead and lead compounds. Industrial hygienists and other public health professionals need to determine the effectiveness of measures taken to control workers' exposure, and this is generally achieved by making workplace air measurements.

This standard was originally published in 2003 as an identical adoption of ISO 8518: 2001 under dual numbering. The first revision of this standard has been brought out in order to align it with the latest version of ISO 8518: 2022 In this revision, following modifications have been done

a) A new Annex B (informative) has been added concerning sampler wall deposits;
b) References and definitions have been updated;
c) Additional editorial changes have been made.

This document specifies flame and electrothermal atomic absorption spectrometric methods for the determination of the time-weighted average mass concentration of particulate lead and lead compounds in workplace air.

The text of ISO Standard has been approved as suitable for publication as an Indian Standard without deviations. Certain conventions and terminologies are, however, not identical to those used in Indian Standards. Attention is particularly drawn to the following:

a) Wherever the words `International Standard' appear referring to this standard, they should be read as `IndianStandard'.

b) Comma (,) has been used as a decimal marker in the International Standard, while in Indian Standards, the currentpractice is to use a point (.) as the decimal marker.

In this adopted standard, reference appears to certain International Standards for which Indian Standards also exist. The corresponding Indian Standards, which are to be substituted in their places, are listed below along with their degree of equivalence for the editions indicated:

International Standard	*Corresponding Indian Standard*	*Degree of Equivalence*
ISO 3585, Borosilicate glass 3.3 — Properties	IS 18219 : 2023/ISO 3585:1998 — Borosilicate glass 3.3 - Properties	Identical with ISO 3585:1998
ISO 8655-2, Piston-operated volumetric apparatus — Part 2: Pipettes	IS 17094 (Part 2) : 2019/ ISO 8655-2 : 2002 —Piston - Operated volumetric apparatus: Part 2 piston pipettes	Identical with ISO 8655-2:2002

The technical committee has reviewed the provisions of the following International Standards referred in this adoptedstandard and has decided that they are acceptable for use in conjunction with this standard.

International Standard No.	*Title*
ISO 3696:1987	Water for analytical laboratory use — Specification and test methods
ISO 7708:1995	Air quality — Particle size fraction definitions for health-related sampling
ISO 8655-1	Piston-operated volumetric apparatus — Part 1: Terminology, general requirements and user recommendations
ISO 8655-5	Piston-operated volumetric apparatus — Part 5: Dispensers
ISO 8655-6	Piston-operated volumetric apparatus — Part 6: Gravimetric reference measurement procedure for the determination of volume
ISO 13137	Workplace atmospheres — Pumps for personal sampling of chemical agents — Requirements and test methods

In this adopted standard, reference appears to certain International Standards where the standard atmospheric conditions to be observed are stipulated which are not applicable to tropical/subtropical countries. The applicable standard atmospheric conditions for Indian conditions are (27 ± 2) ℃ and (65 ± 5) percent, relative humidity and shall be observed while using this standard.

In reporting the result of a test or analysis made in accordance with this standard, if the final value, observed or calculated, is to be rounded off, it shall be done in accordance with IS 2 : 2022 'Rules for rounding off numerical values (*second revision*)'.

INTERNATIONAL STANDARD

ISO 8518

Third edition
2022-10

Workplace air — Determination of particulate lead and lead compounds — Flame and electrothermal atomic absorption spectrometric methods

Air des lieux de travail — Dosage du plomb particulaire et des composés particulaires du plomb — Méthode par spectrométrie d'absorption atomique dans la flamme et méthode par spectrométrie d'absorption avec atomisation électrothermique

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Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular, the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. ISO shall not be held responsible for identifying any or all such patent rights. Details of any patent rights identified during the development of the document will be in the Introduction and/or on the ISO list of patent declarations received (see www.iso.org/patents).

Any trade name used in this document is information given for the convenience of users and does not constitute an endorsement.

For an explanation of the voluntary nature of standards, the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the World Trade Organization (WTO) principles in the Technical Barriers to Trade (TBT), see www.iso.org/iso/foreword.html.

This document was prepared by Technical Committee ISO/TC 146, *Air quality*, Subcommittee SC 2, *Workplace atmospheres*.

This third edition cancels and replaces the second edition (ISO 8518:2001), which has been technically revised.

The main changes are as follows:

— a new Annex B (informative) has been added concerning sampler wall deposits;

— references and definitions have been updated;

— additional editorial changes have been made.

Any feedback or questions on this document should be directed to the user's national standards body. A complete listing of these bodies can be found at www.iso.org/members.html.

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Introduction

The health of workers in many industries, for example, mining, metal refining, battery manufacture, construction, is at risk through exposure by inhalation of particulate lead and lead compounds. Industrial hygienists and other public health professionals need to determine the effectiveness of measures taken to control workers' exposure, and this is generally achieved by making workplace air measurements. This document provides a method for making valid exposure measurements for lead. It will be of benefit to:

— agencies concerned with health and safety at work;

— industrial hygienists and other public health professionals;

— analytical laboratories;

— industrial users and workers of metals and metalloids, etc.

During the development of this document, it has been assumed that the execution of its provisions and the interpretation of the results obtained is entrusted to appropriately qualified and experienced people.

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Workplace air — Determination of particulate lead and lead compounds — Flame and electrothermal atomic absorption spectrometric methods

1 Scope

This document specifies flame and electrothermal atomic absorption spectrometric methods for the determination of the time-weighted average mass concentration of particulate lead and lead compounds in workplace air.

These methods are typically applicable to personal sampling of the inhalable fraction of airborne particles, as defined in ISO 7708, and to static (area) sampling. It can be applied to other health-related fractions as required.

The sample dissolution procedure specifies hot plate or microwave assisted digestion, or ultrasonic extraction (see 11.2). The use of an alternative, more vigorous dissolution procedure is necessary when it is desired to extract lead from compounds present in the test atmosphere that are insoluble using the dissolution procedures described herein (see Clause 5).

The flame atomic absorption method is applicable to the determination of masses of approximately 1 µg to 200 µg of lead per sample, without dilution[1]. The electrothermal atomic absorption method is applicable to the determination of masses of approximately 0,01 µg to 0,5 µg of lead per sample, without dilution[1].

The ultrasonic extraction procedure has been validated for the determination of masses of approximately 20 µg to 100 µg of lead per sample, for laboratory-generated lead fume air filter samples[2].

The concentration range for lead in air for which this procedure is applicable is determined in part by the sampling procedure selected by the user (see 10.1).

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2 Normative references

The following documents are referred to in the text in such a way that some or all of their content constitutes requirements of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 3585, *Borosilicate glass 3.3 — Properties*

ISO 3696:1987, *Water for analytical laboratory use — Specification and test methods*

ISO 7708:1995, *Air quality — Particle size fraction definitions for health-related sampling*

ISO 8655-1, *Piston-operated volumetric apparatus — Part 1: Terminology, general requirements and user recommendations*

ISO 8655-2, *Piston-operated volumetric apparatus — Part 2: Pipettes*

ISO 8655-5, *Piston-operated volumetric apparatus — Part 5: Dispensers*

ISO 8655-6, *Piston-operated volumetric apparatus — Part 6: Gravimetric reference measurement procedure for the determination of volume*

ISO 13137, *Workplace atmospheres — Pumps for personal sampling of chemical agents — Requirements and test methods*

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ISO 15202-2, *Workplace air — Determination of metals and metalloids in airborne particulate matter by inductively coupled plasma atomic emission spectrometry — Part 2: Sample preparation*

ISO/IEC 17025, *General requirements for the competence of testing and calibration laboratories*

ISO 17034, *General requirements for the competence of reference material producers*

ISO 18158, *Workplace air — Terminology*

ISO 20581, *Workplace air — General requirements for the performance of procedures for the measurement of chemical agents*

EN 13205, *Workplace atmospheres — Assessment of performance of instruments for measurement of airborne particle concentrations*

3 Terms and definitions

For the purposes of this document, the terms and definitions given in ISO 18158 and the following apply.

ISO and IEC maintain terminology databases for use in standardization at the following addresses:

— ISO Online browsing platform: available at https://www.iso.org/obp

— IEC Electropedia: available at https://www.electropedia.org/

3.1
sample dissolution
process of obtaining a solution containing all analytes of interest from a sample, which might or might not involve complete dissolution of the sample

[SOURCE: ISO 15202-2:2020, 3.1]

3.2
sample solution
solution prepared from a sample by the process of *sample dissolution* (3.1)

Note 1 to entry: A sample solution might need to be subjected to further operations, e.g. dilution, or addition, or both, of an internal standard(s), in order to produce a *test solution* (3.3).

[SOURCE: ISO 15202-2:2020, 3.2]

3.3
test solution
blank solution or *sample solution* (3.2) that has been subjected to all operations required to bring it into a state in which it is ready for analysis

[SOURCE: ISO 15202-2:2020, 3.3, modified — Note 1 to entry has been deleted.]

4 Principle

4.1 A known volume of air is drawn through a sampling substrate to collect particulate lead and lead compounds. For personal sampling, a sampler designed to collect the inhalable fraction of airborne particles is typically used.

4.2 The sampling substrate and collected sample are subjected to a dissolution procedure in order to extract lead. The sample dissolution procedure can use one of three techniques: hot plate digestion, microwave assisted digestion or ultrasonic extraction.

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4.3 Sample solutions are analysed for lead content by aspirating into the oxidizing air-acetylene flame of an atomic absorption spectrometer equipped with a lead hollow-cathode lamp or electrodeless discharge lamp. Absorbance measurements are made at 283,3 nm with background correction (e. g. Zeeman mode or deuterium background correction), and analytical results are obtained by the analytical curve technique. Potential interference by anions that form precipitates with lead is overcome by the addition of the disodium salt of ethylenediamine tetraacetic acid (EDTA) when necessary.

4.4 For accurate lead determination when the concentration of lead in the solution is low, the analysis can be repeated using electrothermal atomic absorption spectrometry. Aliquots of the test solution are injected into a graphite furnace, and after drying and sample ashing stages, the sample is atomized electrothermally. Absorbance measurements are made at 283,3 nm with background correction and results are obtained by the analytical curve technique.

4.5 The results can be used for the assessment of workplace exposures to airborne particulate lead (see ISO 21832[3]).

5 Reactions

In general, the overwhelming majority of particulate lead compounds that are commonly found in samples of workplace air are converted to water-soluble lead ions (Pb^{2+}) by the sample dissolution procedures described in 11.2. However, certain lead compounds, for example, lead-containing silicates, can possibly not be dissolved. If necessary, a dissolution procedure employing hydrofluoric acid should be used to dissolve silicate lead. If there is any doubt about the effectiveness of these procedures for the dissolution of particulate lead compounds that can be present in the test atmosphere, then this shall be investigated before proceeding with the analytical method described in Clause 11.

6 Requirement

The measuring procedure shall comply with applicable requirements of ISO 20581 and with any relevant national standard that specifies performance requirements for procedures for measuring chemical agents in workplace air.

7 Reagents

During the analysis, use only reagents of recognized analytical grade, and only water as specified in 7.1.

7.1 Water, conforming with the requirements for ISO 3696:1987, grade 2 water (electrical conductivity less than 0,1 mS/m and resistivity greater than 0,01 MΩ·m at 25 °C).

The concentration of lead in the water shall be less than 0,01 µg/ml.

It is recommended that the water used be obtained from a water purification system that delivers ultrapure water having a resistivity greater than 0,18 MΩ·m (usually expressed by manufacturers of water purification systems as 18 MΩ·cm).

7.2 Nitric acid (HNO_3), concentrated, $\rho \approx 1{,}42$ g/ml (about 70 % mass fraction).

The concentration of lead shall be less than 0,01 µg/ml.

WARNING — Concentrated nitric acid is corrosive and oxidizing, and nitric acid fumes are irritant. Avoid exposure by contact with the skin or eyes, or by inhalation of fumes. Use suitable personal protective equipment (including suitable gloves, face shield or safety glasses, etc.) when working with the concentrated or dilute nitric acid, and carry out sample dissolution with concentrated nitric acid in open vessels in a fume hood.

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7.3 Nitric acid, diluted 1 + 1.

Carefully add 500 ml of concentrated nitric acid (7.2) to 450 ml of water (7.1) in a 2-litre beaker. Swirl to mix, allow to cool and transfer to a 1-litre one-mark volumetric flask. Dilute to the mark with water, stopper and mix thoroughly.

7.4 Nitric acid, diluted 1 + 9.

Place approximately 800 ml of water (7.1) in a 1-litre one-mark volumetric flask. Carefully add 100 ml of concentrated nitric acid (7.2) to the flask and swirl to mix. Allow to cool, dilute to 1 litre with water and mix thoroughly.

7.5 Hydrofluoric acid (HF), concentrated, with the density, ρ, almost equal to 1,14 g/ml (about 38 % mass fraction), if required, for digestion of samples containing lead silicates.

The concentration of lead in the HF shall be less than 0,1 µg/ml.

WARNING — Concentrated hydrofluoric acid and hydrogen fluoride vapour are extremely toxic and intensely corrosive, and diluted hydrofluoric acid can also cause serious and painful burns that can possibly not be felt until up to 24 h after contact. Avoid exposure by contact with the skin or the eyes, or by inhalation of the vapour. Use of personal protection (for example, impermeable gloves, face shield or safety glasses) is essential when working with concentrated or diluted hydrofluoric acid, and concentrated hydrofluoric acid should be used in a fume hood. It is essential that hydrofluoric acid antidote gel containing calcium gluconate is readily available to workers, both during and for 24 h after use of hydrofluoric acid.

7.6 Matrix modifier, $NH_4H_2PO_4$, $Mg(NO_3)_2$ or $Pd(NO_3)_2$, or a combination of these, if required, for analysis by electrothermal atomic absorption spectrometry.

7.7 Stock lead standard solution, 1 000 mg/l of lead.

Use a commercial standard solution with a certified lead concentration traceable to national standards. Observe the manufacturer's expiration date or recommended shelf life.

Alternatively, prepare a lead standard solution by one of the following procedures.

a) Dissolve 1,598 g ± 0,001 g of lead(II) nitrate [$Pb(NO_3)_2$], previously dried to constant mass at 110 °C and cooled in a desiccator, in 200 ml of 1 + 1 nitric acid (7.3). Quantitatively transfer the solution to a 1 000 ml one-mark volumetric flask. Dilute to the mark with water (7.1), stopper and mix thoroughly. Store in a suitable container, for example, a polypropylene bottle (8.6.2.2), for a maximum period of one year.

b) Dissolve 1,000 g ± 0,001 g of lead wire (99,9 % mass fraction Pb) in 200 ml of 1 + 1 nitric acid (7.3). Quantitatively transfer the solution into a 1 000 ml one-mark volumetric flask, dilute to the mark with water (7.1), stopper and mix thoroughly. Store in a suitable container, for example, a polypropylene bottle (8.6.2.2), for a maximum period of one year.

7.8 Working lead standard solution, 1 mg/l of lead, if required, for analysis by electrothermal atomic absorption spectrometry.

Accurately pipette 100 µl of stock lead standard solution (7.7) into a 100 ml one-mark volumetric flask (8.6.1.4). Add 1 ml of concentrated nitric acid (7.2), dilute to the mark with water (7.1), stopper and mix thoroughly. Store in a suitable container, for example, a polypropylene bottle (8.6.2.2), for a maximum period of one month.

7.9 Hydrogen peroxide (H_2O_2), approximately 30 % mass fraction solution, if required, for use in the hot-plate sample digestion method.

The concentration of lead in the hydrogen peroxide solution shall be less than 0,01 µg/ml.

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7.10 Acetylene, if required, for use in analysis by flame atomic absorption spectrometry.

7.11 Air, compressed and filtered, if required, for use in analysis by flame atomic absorption spectrometry.

8 Apparatus

8.1 Inhalable samplers, designed to collect the inhalable fraction of airborne particles, complying with the provisions of EN 13205, for use when the exposure limits of interest apply to the inhalable fraction of airborne particles.

NOTE 1 In general, personal samplers for collection of the inhalable fraction of airborne particles do not exhibit the same size selective characteristics if used for static (area) sampling.

NOTE 2 Some inhalable samplers are designed to collect the fraction of airborne particles on a sampling substrate, and any particulate matter deposited on the internal surfaces of the sampler is not of interest. Other inhalable samplers are designed such that airborne particles that pass through the entry orifice(s) match the inhalable convention, in which case particulate matter deposited on the internal surfaces of the sampler does form part of the sample. (Samplers of this second type generally incorporate an internal filter cassette or cartridge that can be removed from the sampler to enable this material to be easily recovered.) The operating instructions supplied by the manufacturer should be consulted to find out whether particulate matter deposited on the internal surfaces of the sampler forms part of the sample. Annex B provides additional guidance on sampler wall deposits.

NOTE 3 Samplers to collect other fractions defined in ISO 7708 can be used when necessary (for example, when there are occupational exposure limit values associated with those fractions.)

8.2 Sampling substrate, (for example, a filter) of a diameter suitable for use with the samplers (see 8.1), with a collection efficiency of not less than 99,5 % for particles with a 0,3 µm diffusion diameter in accordance with 2.2 of ISO 7708:1995, with a maximum lead content (typically less than 0,1 µg Pb), and compatible with the selected sample preparation method.

NOTE 1 Guidance on filter selection is provided in Annex A.

NOTE 2 Digestible cellulosic capsules (consisting of cellulose acetate housing attached to cellulosic filter) that are placed within certain types of samplers are now available for use, thereby accounting for potential internal aerosol wall losses[4],[5].

8.3 Sampling pumps, meeting the specifications of ISO 13137.

8.4 Flowmeter, portable, with an accuracy that is sufficient to enable the volumetric flow rate (see 10.1.1.2) to be measured to within ±5 %.

The calibration of the flowmeter shall be checked against a primary standard, i.e. a flowmeter whose accuracy is traceable to national standards. If appropriate (see 10.1.3.1), record the atmospheric temperature and pressure at which the calibration of the flowmeter was checked.

8.5 Ancillary equipment

8.5.1 Flexible tubing, of a diameter suitable for making a leak-proof connection from the samplers to the sampling pumps.

8.5.2 Belts or harnesses, to which the sampling pumps can conveniently be fixed for personal sampling (except where the sampling pumps are small enough to fit inside worker's pockets).

8.5.3 Flat-tipped forceps, for loading and unloading sampling substrates into samplers.

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8.5.4 Transport cassettes, or similar, if required to transport samples for laboratory analysis.

8.5.5 Barometer, (readable to 0,1 kPa), suitable for measurement of atmospheric pressure, if required (see 10.1.3).

8.5.6 Thermometer, (readable to 1 °C), minimum temperature range of 0 °C to 50 °C, with graduated divisions of 1 °C or less, for measurement of atmospheric temperature.

For applications at temperatures below freezing, the range of the thermometer shall extend to the appropriate desired range.

8.6 Analytical or laboratory apparatus

Ordinary laboratory apparatus, and the following.

8.6.1 Glassware, made of borosilicate glass 3.3 and conforming with the requirements of ISO 3585.

It is preferable to reserve a set of glassware for analysis of lead by this method, in order to ensure that problems do not arise from incomplete removal of lead contamination by cleaning.

8.6.2 Plastic labware, including the following:

8.6.2.1 Heatable beakers, beaker covers, etc., if required, made of a material that is resistant to corrosion by hydrofluoric acid, for example, a fluorocarbon polymer such as polytetrafluoroethylene (PTFE), and suitable for performing dissolutions using hydrofluoric acid.

8.6.2.2 Polypropylene bottles, of capacities from 100 ml to 1 000 ml.

8.6.3 Piston-operated volumetric instruments, complying with the requirements of ISO 8655-1 and tested in accordance with ISO 8655-6:

8.6.3.1 Pipetters, complying with the requirements of ISO 8655-2, as an alternative to one-mark pipettes, for the preparation of standard solutions, calibration solutions and dilution of samples.

8.6.3.2 Dispensers, complying with the requirements of ISO 8655-5, for dispensing acids.

8.6.4 Hot plate, thermostatically controlled, capable of maintaining a surface temperature of approximately 150 °C, for hot-plate procedures.

8.6.5 Microwave digestion apparatus

8.6.5.1 General

Ensure that manufacturer's safety recommendations are followed.

NOTE 1 The specified method is for closed vessel microwave digestion systems with a temperature control system. Microwave digestion systems that are equipped only with either a pressure control system or lower pressure vessels, or both, can be used provided that a suitable sample dissolution procedure is developed, and a prior assessment of dissolution efficiency is carried out.

NOTE 2 Open-vessel microwave digestion systems can give results equivalent to closed-vessel microwave digestion systems. They can therefore be used provided that a suitable sample dissolution procedure is developed, and a prior assessment of dissolution efficiency is carried out.

8.6.5.2 Microwave digestion system, designed for closed-vessel sample digestion in the laboratory, with power output regulation, fitted with a temperature control system capable of minimum sensing

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the temperature to within ±2 °C and automatically adjusting the microwave power output within minimum 2 s.

The microwave cavity shall be corrosion-resistant and well ventilated, with all electronics protected against corrosion to ensure safe operation.

CAUTION — Do not use domestic (kitchen) microwave ovens, since there are very significant hazards associated with their use for the procedure described in this document. Acid vapours released into the cavity can corrode safety devices that prevent the magnetron from shutting off when the door is opened, potentially exposing the operator to microwave energy. Also, the fumes generated can be extremely hazardous.

NOTE A pressure control system is also very useful, since it provides a safeguard against the possibility of sample loss due to excessive pressure build-up and partial venting of the sample vessels.

8.6.5.3 Vessels, designed for carrying out microwave assisted digestions, capable of withstanding a temperature of 180 °C, and with an internal volume of at least 50 ml.

The vessels shall be transparent to microwave energy, and shall be capable of withstanding internal pressures up to at least 3 000 kPa or greater, and temperatures up to at least 180 °C or greater.

Closed vessels shall also be equipped with a safety relief valve or disc that will prevent vessel rupture or ejection of the vessel cap. Such vessels consist of an inner liner and cover made of a microwave-transparent and chemically resistant material [usually a fluorocarbon polymer such as tetrafluoromethoxyl polymer (TFM)], which contains and isolates the sample solution from a high-strength, outer pressure vessel structure. Other types of sample vessel designed to operate at equivalent or higher temperatures or pressures, or both, can be used. If hydrofluoric acid (HF) is to be used, the vessels shall be compatible with HF.

CAUTION — For closed-vessel designs, the material from which the outer vessels are made is usually not as chemically resistant as the liner material. Since the outer vessels provide the strength required to withstand the high pressures within the inner liners, they shall be inspected regularly to check for any chemical or physical degradation.

NOTE Alternatively, a one reaction chamber microwave assisted pressure digestion system can be used.

8.6.6 Ultrasonic bath (sonicator), for performing ultrasonic extractions, capable of delivering sufficient power to effect the quantitative dissolution of particulate lead under the conditions described in 11.2.5 (typically 1 W/cm^2 power density or greater).

8.6.7 Plastic centrifuge tubes, 50 ml, with screw caps (for ultrasonic procedure).

8.6.8 Atomic absorption spectrometer, fitted with an air-acetylene burner supplied with compressed air and acetylene, and equipped with either a lead hollow cathode lamp or electrodeless discharge lamp[6],[7]. If sample dissolution is carried out with the aid of hydrofluoric acid (see 11.2.3.3 and 11.2.4.2), the atomic absorption spectrometer shall be hydrofluoric acid-compatible. If electrothermal atomic absorption is to be carried out, the atomic absorption spectrometer shall be capable of carrying out simultaneous background correction at 283,3 nm, either by using a continuum source such as a deuterium lamp to measure non-specific attenuation, or by using Zeeman or Smith-Hieftje background correction systems[8].

8.6.9 Electrothermal atomiser, fitted with a solid, pyrolytic graphite platform mounted in a pyrolytically-coated graphite tube, supplied with argon purge gas, and equipped with an autosampler capable of injecting microlitre volumes onto the platform.

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NOTE Some manufacturers of atomic absorption spectrometers use an alternative design of electrothermal atomiser to achieve a constant temperature environment during atomisation, and some use aerosol deposition as a means of sample introduction. The use of such accessories is acceptable, provided satisfactory method performance is verified. Likewise, atomisers made from heat-resistant metal, such as tungsten, can also be suitable.

8.6.10 Analytical balance, capable of weighing to ±0,1 mg, if required, for use in preparation of stock standard lead solution.

8.6.11 Disposable gloves, for prevention of sample contamination.

9 Occupational exposure assessment

9.1 Assessment strategy

Refer to relevant International or national Standards (for example, EN 689[9] and ASTM E1370[10]) for guidance on how to develop an appropriate assessment strategy.

9.2 Measurement strategy

9.2.1 General

Refer to relevant International or national Standards (for example, EN 689[9] and ASTM E1370[10]) for general guidance on measurement strategy.

9.2.2 Personal sampling

Exposure of workers to lead shall normally be determined by personal sampling, since the concentration of lead and lead compounds in the breathing zone is usually higher than their background levels in the workplace.

9.2.3 Static (area) sampling

Static (area) sampling can be carried out, if appropriate, to assess the exposure of workers in a situation where personal sampling is not possible; to characterise the background level of lead in the workplace to give an indication of the efficiency of ventilation or other engineering controls; or to provide information on the location and intensity of an emission source.

9.3 Selection of measurement conditions and measurement pattern

9.3.1 General

9.3.1.1 The sampling procedure shall be devised to provide samples that are representative of normal working conditions and that are compatible with the analytical method while causing the least possible interference with the worker and the normal performance of the job.

9.3.1.2 The pattern of sampling shall take into consideration practical issues, such as the nature of the measurement task and the frequency and duration of particular work activities.

NOTE Additional information concerning purposes of measurement and performance requirements can be found in ISO 20581.

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9.3.2 Screening measurements of time-weighted average concentration and worst-case measurements

Screening measurements of time-weighted average concentration can be carried out in the initial stages of a survey to assess the effectiveness of control measures. This can involve sampling during representative work episodes to obtain clear information about the level and pattern of exposure, or worst-case measurements can be made.

9.3.3 Screening measurements of variation of concentration in either time or space, or both

Screening measurements of variation of concentration in either time or space, or both, can be carried out in the initial stages of a survey to identify locations and periods of elevated exposure, and to set the duration and frequency of sampling for measurements for comparison with limit values.

NOTE For making screening measurements of variation of concentration in either time or space, or both, the sampling time used is normally between 5 min and 30 min.

9.3.4 Measurements for comparison with limit values and periodic measurements

For making long-term measurements, samples shall be collected for the entire working period or during a number of representative work episodes[11].

NOTE The best estimate of long-term exposure is obtained by taking samples for the entire working period, but this is often not practicable or not desirable (for example, because of the possibility of overloading the sampling substrate).

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10 Sampling

10.1 Preliminary considerations

10.1.1 Selection and use of samplers

10.1.1.1 Select samplers as described in 8.1.

If possible, the samplers selected should be manufactured from conducting material, since samplers composed of non-conducting material have electrostatic properties that can influence representative sampling.

10.1.1.2 Use the samplers at their design flow rate and in accordance with the manufacturer's instructions.

10.1.2 Sampling period

10.1.2.1 Select a sampling period long enough to ensure that the amount of lead collected is adequate to enable lead-in-air concentrations to be determined at the required level (see 9.3).

10.1.2.2 In calculating the minimum sampling time required it is necessary to consider the selected flow rate and the lower limit of the analytical working range of the method[12].

10.1.2.3 When high concentrations of airborne particles are anticipated, select a sampling period that is not so long as to risk overloading the sampling substrate with particulate matter.

NOTE If overloading is observed or suspected problem and it is desired to sample for the entire working day, it can be necessary to collect consecutive samples[13].

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10.1.3 Temperature and pressure effects

10.1.3.1 Expression of results

Consider whether it is necessary to recalculate the concentration of lead in air to reference conditions (such as in high altitude situations). If so, measure and record the atmospheric temperature and pressure at the start and at the end of the sampling period (see 10.4.1 and 10.4.2) and use the formula given in C.2 to apply the necessary correction.

NOTE The concentration of lead in air is generally stated for actual environmental conditions (temperature, pressure) at the workplace during the sampling period.

10.1.3.2 Effect of temperature and pressure on flow rate measurements

Refer to the manufacturer's instructions to determine if the indicated volumetric flow rate of the flowmeter (8.4) is dependent upon temperature and pressure. Consider whether the difference between the atmospheric temperature and pressure at the time of calibration of the flowmeter and during sampling is likely to be great enough to justify making a correction to take this into account, for example, if the error can be greater than ±5 %. If a correction is necessary, measure and record the atmospheric temperature and pressure at which the calibration of the flowmeter was checked (see 8.4) and measure and record the atmospheric temperature and pressure at the start and at the end of the sampling period (see 10.4.1 and 10.4.2).

NOTE An example of temperature and pressure correction for the indicated volumetric flow rate is given in C.1 for a constant pressure drop, variable area, flowmeter.

10.2 Preparation of sampling equipment

10.2.1 Cleaning of samplers

Unless disposable cassettes are used, clean the samplers (8.1) before use. Disassemble the samplers, soak in detergent solution, rinse thoroughly with water, wipe with absorbent tissue and allow to dry before reassembly. Alternatively, use a laboratory washing machine.

NOTE If necessary, ultrasound can be used to remove fine dust.

10.2.2 Loading the samplers with sampling substrate

Using flat-tipped forceps (8.5.3), load clean samplers (see 10.2.1) with sampling substrate (8.2), label each sampler so that it can be uniquely identified and seal with its protective cover or plug to prevent contamination.

Alternatively, commercially available pre-loaded cassettes can be used.

10.2.3 Setting the volumetric flow rate

Perform the following in a clean area, where the concentration of lead is low:

Connect each loaded sampler (see 10.2.2) to a sampling pump (8.3) using flexible tubing (8.5.1), ensuring that no leaks can occur. Remove the protective cover or plug from each sampler, switch on the sampling pump, attach the flowmeter (8.4) to the sampler so that it measures the flow through the sampler inlet orifice(s), and set the required volumetric flow rate (see 10.1.1.2). Switch off the sampling pump and seal the sampler with its protective cover or plug to prevent contamination during transport to the sampling position.

If necessary, allow the sampling pump operating conditions to stabilize before setting the volumetric flow rate.

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10.2.4 Field blanks

Retain as field blanks one unused loaded sampler from each batch of ten prepared, subject to a minimum of three. Treat these in the same manner as those used for sampling in respect of storage and transport to and from the sampling position but draw no air through the sampling substrate.

10.3 Sampling position

10.3.1 Personal sampling

Position the sampler in the worker's breathing zone, as close to the mouth and nose as is reasonably practicable, for example, fastened to the worker's lapel. Attach the sampling pump to the worker in a manner that causes minimum inconvenience, for example, to a belt (8.5.2) around the waist or place it in a convenient pocket.

10.3.2 Static (area) sampling

10.3.2.1 If static sampling is carried out to assess the exposure of a worker in a situation where personal sampling is not possible (for example, due to the need to sample at a volumetric flow rate higher than the design flow rate of available personal samplers), position the sampler in the immediate vicinity of the worker and at breathing height. If in doubt, take the sampling position to be the point where the risk of exposure is considered to be greatest.

10.3.2.2 If static sampling is carried out to characterize the background level of lead in the workplace, select a sampling position that is sufficiently remote from the work processes, such that results will not be directly affected by lead from emission sources.

10.4 Collection of samples

10.4.1 When ready to begin sampling, remove the protective cover or plug from the sampler and switch on the sampling pump. Record the time and volumetric flow rate at the start of the sampling period. If the sampling pump is fitted with an integral timer, check that this is reset to zero. If appropriate (see 10.1.1.2), measure the atmospheric temperature and pressure at the start of the sampling period using the thermometer (8.5.6) and barometer (8.5.5), and record the measured values.

NOTE If the temperature or pressure at the sampling position is different from that where the volumetric flow rate was set (see 10.2.3), the volumetric flow rate can change, and it can need re-adjusting before sampling.

10.4.2 At the end of the sampling period (see 10.1.2), switch off the pump, record the time and calculate the duration of the sampling period. Check either the malfunction indicator or the reading on the integral timer, or both, if fitted, and consider the sample to be invalid if there is evidence that the sampling pump was not operating properly throughout the sampling period. Measure the volumetric flow rate at the end of the sampling period using the flowmeter (8.4) and record the measured value. If appropriate (see 10.1.3), measure the atmospheric temperature and pressure at the end of the sampling period using the thermometer (8.5.6) and barometer (8.5.5), and record the measured values.

10.4.3 Carefully record the sample identity and all relevant sampling data (see Clause 14). Calculate the mean volumetric flow rate by averaging the volumetric flow rates at the start and at the end of the sampling period and, if appropriate (see 10.1.3), calculate the mean atmospheric temperature and pressure. Calculate the volume of air sampled, in litres, at atmospheric temperature and pressure, by multiplying the mean flow rate in litres per minute by the duration of the sampling period in minutes.

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10.5 Transportation

10.5.1 For samplers which collect airborne particles on the sampling substrate (see Note 2 in 8.1), remove the sampling substrate from each sampler, place in a labelled transport cassette (8.5.4) and close with a lid. Take particular care to prevent the collected sample from becoming dislodged from heavily loaded sampling substrate. Alternatively, transport samples to the laboratory in the samplers in which they were collected.

10.5.2 For samplers which have an internal cassette (see Note 2 in 8.1), carefully remove the cassette from each sampler and fasten with its lid or transport clip.

10.5.3 For samplers of the disposable cassette type, transport samples to the laboratory in the samplers in which they were collected.

10.5.4 Transport the samples (10.5.1 to 10.5.3) to the laboratory in a container that has been designed to prevent damage to the samples in transit and which has been labelled to assure proper handling.

10.5.5 Follow sampling chain of custody procedures to ensure sample traceability. Ensure that the documentation which accompanies the samples is suitable for a "chain of custody" to be established (see, for example, ASTM D4840[14]).

10.6 Storage

In the laboratory, the sealed samplers can be stored at room temperature and normal humidity. Losses of sample can occur, if pressure is applied to the surface of the dust collected on a sampling substrate, especially during the transfer of the substrate from a capsule. For example, sample losses can occur if the sample surface comes into contact with tweezers or the edge of the sampler.

NOTE Sample substrates can become charged during sampling and can attract themselves to these items. Losses of dust from the surface or found in the capsule shall be noted on the report.

11 Analysis

11.1 Cleaning of glassware and plasticware

11.1.1 Perform all of the following while wearing appropriate personal protective equipment (for example, gloves, safety glasses).

11.1.2 Before use, clean all glassware, microwave digestion vessels, and plasticware to remove any residual grease or chemicals by first soaking in laboratory detergent solution and then rinsing thoroughly with water (7.1).

11.1.3 After initial cleaning with detergent and water, clean all beakers with nitric acid. This can be accomplished either by soaking for a minimum of 24 h in concentrated nitric acid (7.2), or by the following procedure. Fill beakers to one-third capacity with concentrated nitric acid (7.2), and then heat them on a hot plate with a surface temperature of 140 °C in a fume hood until most of the liquid has evaporated and allow to cool. Rinse beakers thoroughly with water (7.1).

11.1.4 Glassware that has been previously subjected to the entire cleaning procedure described in the previous steps, and which has been reserved for the analysis of lead, can be cleaned adequately by rinsing with 1 + 9 nitric acid (7.4) and then with water (7.1).

11.1.5 Before use, clean polypropylene bottles, microwave digestion vessels and other plasticware by soaking them in 1 + 9 nitric acid (7.4) for at least 24 h and then rinse thoroughly with water (7.1).

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NOTE 1 Plasticware (possibly disposable) can be received in clean condition directly from the vendor, thereby precluding the need for cleaning prior to use.

NOTE 2 Alternatively a laboratory automatic washer (with acid rinse cycle) can be used.

11.2 Preparation of sample and blank solutions

11.2.1 General

Perform all of the following preparations while wearing appropriate personal protective equipment (for example, gloves, safety glasses).

11.2.2 Selection of sample dissolution method

Prepare samples and blanks for analysis using one of the three sample preparation methods described below: either hot-plate digestion, microwave assisted digestion or ultrasonic extraction.

11.2.3 Hot plate digestion method

11.2.3.1 Open the samplers, sampler cassettes or transport cassettes (see 10.5), and transfer each sample or blank into a clean, labelled 50 ml beaker (8.6.1.1) using flat-tipped forceps (8.5.3). If the sampler used was of a type in which airborne particles deposited on the internal surfaces of the sampler form part of the sample, wash any particulate matter adhering to the internal surfaces into the beaker using a minimum volume of 1 + 9 nitric acid (7.4).

11.2.3.2 To each beaker, add 3 ml of concentrated nitric acid (7.2) and 1 ml of hydrogen peroxide (7.9), and cover with a watch-glass.

11.2.3.3 Heat on a hot plate (8.6.4) with a surface temperature of approximately 140 °C in a fume hood and allow the solution to evaporate until the final solution volume is reduced to approximately 1 ml. Avoid taking to dryness. Remove beakers from the hot plate and allow to cool.

NOTE The exact hot-plate temperature is not critical. A temperature of 140 °C is used because it is high enough to enable the liquid to be evaporated at an acceptable rate. This temperature is also useful for minimizing the risk of taking samples to dryness.

The use of hydrofluoric acid (HF) (7.5) is required to dissolve silicate lead. If the material in the test atmosphere is believed to contain a significant amount of silicate material, its dissolution can be facilitated by adding 1 ml of HF at the same time as the nitric acid. However, it will be necessary to use heatable beakers and beaker covers, etc. (8.6.2.1), that are made of plastic that is resistant to corrosion by HF, for example, a fluorocarbon polymer such as PTFE.

11.2.3.4 Carefully rinse each watch-glass and the sides of each beaker with water and transfer each solution quantitatively to a 10 ml one-mark volumetric flask. If necessary, remove any undissolved particulate by filtration or centrifugation. Dilute to the mark of the volumetric flask with water (7.1), seal the flask with a stopper, and mix thoroughly.

11.2.4 Microwave assisted digestion method

11.2.4.1 Open the samplers, sampler cassettes or transport cassettes (see 10.5), and transfer each sampling substrate into the clean liner of a labelled microwave digestion vessel (8.6.5.2) using flat-tipped forceps (8.5.3). Follow the same procedure for blank sampling substrate. If the sampler used was of a type in which airborne particles deposited on the internal surfaces of the sampler form part of the sample, wash any particulate matter adhering to the internal surfaces into the vessel liner using a minimum volume of water (7.1).

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11.2.4.2 Carefully add 5 ml of concentrated nitric acid (7.2) to the inside of liner of the microwave digestion vessel containing the sample or blank. Seal the vessels.

The use of hydrofluoric acid (7.5) is required to dissolve lead silicates. If the material present in the test atmosphere is believed to contain a significant amount of silicate material, its dissolution can be facilitated by adding 1 ml of hydrofluoric acid at the same time as the nitric acid.

11.2.4.3 Load the vessels into the microwave oven (8.6.5.1) according to manufacturer's instructions. Vessels containing samples shall be evenly and symmetrically placed in the microwave oven.

Even, symmetrical spacing of vessels is needed to ensure uniform microwave heating of all vessel solutions.

11.2.4.4 Program the microwave digestion system to reach 180 °C in less than 10 min, and then hold at this temperature for 15 min.

If hydrofluoric acid is used to dissolve the samples and the temperature sensor is not resistant to attack by this acid, the vessel in which the temperature sensor is fitted should contain a sampling substrate blank in which an equal volume of nitric acid is substituted for the hydrofluoric acid used for dissolution of the samples.

11.2.4.5 At the end of the digestion period, remove the vessels from the microwave oven, place them in a fume hood, and allow the solutions to cool to room temperature.

11.2.4.6 For closed vessels, carefully detach the vent tubing, and carefully shake the vessels to vent any excess gas pressure that can be present inside the vessels. Carefully open each sample vessel.

11.2.4.7 Quantitatively transfer the contents of each vessel to 10 ml-one-mark volumetric flasks. Carefully rinse each vessel with water and dilute to volume with water (7.1). If necessary, remove any undissolved particles by filtration or centrifugation. Seal each flask with a stopper and mix thoroughly.

11.2.5 Ultrasonic extraction method

11.2.5.1 The following method is not applicable to samples containing silicates. In such a case, use the method for sample dissolution using hydrofluoric and nitric acids and ultrasonic agitation as described in ISO 15202-2.

11.2.5.2 Open the samplers, sampler cassettes or transport cassettes (see 10.5), and transfer each sample or blank into a clean 50 ml centrifuge tube (8.6.7) using flat-tipped forceps (8.5.3). Label each centrifuge tube with a unique identifier. If the sampler used was of a type in which airborne particles deposited on the internal surfaces of the sampler form part of the sample, wash any particulate matter adhering to the internal surfaces into the centrifuge tube using a minimum volume of water (7.1). Using a clean glass or plastic rod, push the sampling substrate to the bottom of the centrifuge tube.

11.2.5.3 Introduce 10 ml of 1 + 9 nitric acid (7.4) into each centrifuge tube containing a sample or sampling substrate blank and cap each tube.

11.2.5.4 Place each centrifuge tube upright in an ultrasonic bath (8.6.6) and ensure that the water level within the bath is at or above the level of liquid within the tube.

NOTE Depending on the size of the ultrasonic bath, many centrifuge tubes can be immersed in the bath at one time. A custom rack for the centrifuge tubes can be purchased or constructed to allow for the regular and orderly placement of multiple tubes in the sonicator bath.

11.2.5.5 Apply ultrasonic energy to the acid-immersed samples for a minimum of 30 min. Avoid applying energy at levels that would cause evaporation of the sample.

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11.2.5.6 Remove centrifuge tubes from the bath. Keep tubes in upright position, and allow to cool to room temperature.

11.3 Instrumental analysis

11.3.1 Selection of analytical line

The 283,3 nm lead analytical line shall be used for making absorbance measurements.

NOTE The most sensitive lead line is at 217,0 nm. However, this line is subject to possible spectral interference from antimony, and the significant spectral background at 217,0 nm makes correction for non-specific attenuation (see 5.1.5 of ISO 6955:1982) essential at this wavelength. The 283,3 nm line exhibits somewhat lower sensitivity than the 217,0 nm line, but it is not subject to spectral interference. In addition, while the detection limits obtained are dependent upon the instrument used, absorbance measurements made at 283,3 nm generally have a better signal-to-noise ratio than those made at 217,0 nm, and hence a better detection limit.

11.3.2 Flame atomic absorption spectrometry

11.3.2.1 Instrument setup

Set up the atomic absorption spectrometer (8.6.8) to make absorbance measurements at 283,3 nm, following the manufacturer's instructions for specific instrument operating parameters. Use a lead hollow cathode lamp or electrodeless discharge lamp and an oxidising air-acetylene flame. Allow an appropriate warm-up period for the source lamp.

11.3.2.2 Preparation of calibration solutions

11.3.2.2.1 Use 1 + 9 nitric acid (7.4) as the solvent blank.

11.3.2.2.2 Prepare at least four calibration solutions, including a blank calibration solution, to cover a suitable concentration range, for example, from 0 µg/ml to 20 µg/ml of lead. Accurately pipette appropriate volumes of stock lead standard solution (7.7) into separate, labelled 100 ml one-mark volumetric flasks. Dilute to the mark with 1 + 1 nitric acid (7.3) for test solutions prepared by the microwave assisted digestion method, or with 1 + 9 nitric acid (7.4) for test solutions prepared by the hot plate or ultrasonic sample digestion methods. Stopper and mix thoroughly. Prepare these calibration solutions fresh daily.

NOTE The concentration range of calibration solutions is given as a guide. The upper limit of the working range is dependent upon which wavelength is used, and it is also governed by instrumental factors that affect sensitivity and the linearity of the calibration. Accordingly, the range of the set of calibration solutions can be varied, but when making any changes, ensure that the response of the spectrometer over the alternative range of concentrations selected is such that it is linear.

11.3.2.3 Calibration measurements

11.3.2.3.1 Adjust the spectrometer zero while aspirating the solvent blank (see 11.3.2.2.1) into the flame. Then aspirate the calibration solutions into the flame and make absorbance measurements for each solution.

Use of an autosampler is recommended, since precision is maximised and the volume of solution consumed is minimised.

11.3.2.3.2 Analyse all blank solutions and calculate the mean concentration of the blank solutions.

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11.3.2.4 Calibration measurements

For instruments controlled by a microprocessor or personal computer, use a suitable algorithm to generate the calibration function. For instruments without this capability, prepare a calibration graph by plotting the absorbance of the calibration solutions versus the concentration of lead (µg/ml) in the respective solutions.

In general, it is best to work within the linear range of the calibration, where absorbance is proportional to the lead concentration in solution.

11.3.2.5 Determination

11.3.2.5.1 Adjust the spectrometer zero while aspirating the solvent blank (see 11.3.2.2.1) into the flame. (Repeat this procedure regularly throughout the determination and readjust the zero if the baseline drifts.) Then aspirate the sample and blank test solutions into the flame and make absorbance measurements for each solution. For instruments controlled by a microprocessor or personal computer, use the calibration function to calculate the concentration of lead in the test solutions, and obtain a direct read-out of the results in concentration units. For instruments without this capability, determine the concentration of lead in the test solutions from the calibration graph.

11.3.2.5.2 Aspirate a mid-range calibration solution after each five to ten test solutions and make an absorbance measurement. If this indicates that the sensitivity has changed by more than ±5 %, take one of the following corrective measures: either use the available software facilities of the microprocessor or personal computer to correct for the sensitivity change ("reslope" facility); or suspend analysis and recalibrate the spectrometer. In either case, reanalyse the test solutions that were analysed during the period in which the sensitivity change occurred.

11.3.2.5.3 If high concentrations of lead are found, dilute the sample test solutions to bring the concentration within the calibration range. Make all dilutions so that the final nitric acid concentration is 1 + 9 and record the dilution factor *DF*.

11.3.2.5.4 Calculate the mean lead concentration of the blank test solution.

11.3.2.5.5 If the concentration of lead in the sample test solutions is less than 0,5 µg/ml, consider repeating the analysis using electrothermal atomic absorption spectrometry, since this technique gives more precise measurements at low concentrations.

11.3.3 Electrothermal atomic absorption spectrometry

11.3.3.1 General

Lead is present ubiquitously in the environment, and therefore it is imperative that strict standards of cleanliness are observed to avoid contamination of labware. This is particularly important when carrying out electrothermal atomic absorption spectrometry since the technique exhibits a very low detection limit. Ensure that all glassware is cleaned thoroughly before use, and autosampler cups are stored in 1 + 9 nitric acid until required.

11.3.3.2 Preparation of working calibration solutions

11.3.3.2.1 Prepare a working calibration solution at a concentration of 2,5 ng/ml of lead. Accurately pipette 250 µl of working lead standard solution (7.8) into a 100 ml one-mark volumetric flask. Dilute to the mark with 1 + 1 nitric acid (7.3) for test solutions prepared by the microwave assisted digestion method, or with 1 + 9 nitric acid (7.4) for test solutions prepared by the hot-plate or ultrasonic sample digestion methods. Stopper and mix thoroughly. Prepare this solution fresh weekly.

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11.3.3.2.2 Prepare a working calibration blank solution following the procedure in the preceding paragraph but omitting the 250 µl of working lead standard solution. Prepare this solution fresh weekly.

11.3.3.2.3 For instruments controlled by a microprocessor or personal computer, use a suitable algorithm to generate the calibration function. For instruments without this capability, prepare a calibration graph by plotting the absorbance of the calibration solutions versus the concentration of lead, in micrograms per millilitre, in the respective solutions.

In general, it is best to work in the linear range of the calibration, where absorbance is proportional to the concentration of lead in solution.

11.3.3.3 Calibration and determination

11.3.3.3.1 Set up the atomic absorption spectrometer (8.6.8) and the electrothermal atomiser (8.6.9) to measure lead at a wavelength of 283,3 nm, using background correction to correct for non-specific attenuation (see for example 5.1.5 of ISO 6955:1982). Follow the manufacturer's instructions for specific operating parameters. Allow a suitable warm-up period for the hollow cathode lamp or equivalent source.

NOTE The operating parameters for electrothermal atomic absorption vary considerably between different instruments, much more so than for flame atomic absorption spectrometry.

11.3.3.3.2 Program the autosampler to prepare matrix-modified calibration solutions, sample test solutions and blank test solutions in situ on a pyrolytic graphite platform mounted in the pyrolytically coated graphite tube of the electrothermal atomizer. Prepare at least four matrix-modified calibration solutions to cover the range 0 ng/ml to 50 ng/ml of lead using the working calibration solution (11.3.3.2.1), the working calibration blank solution (11.3.3.2.2), and matrix modifier (7.6). Also prepare matrix-modified sample and blank test solutions using the unmodified sample and blank solutions and matrix-modifier solution. Matrix-modified calibration and test solutions shall be prepared by an autosampler or manually by means of one-mark volumetric flasks.

NOTE The procedure described above can be varied to accommodate the use of electrothermal atomizers of alternative design.

11.3.3.3.3 Set up the analytical sequence in the microprocessor or personal computer interfaced to the electrothermal atomic absorption spectrometric instrument. Specify an appropriate number of replicate analyses for each solution and insert a calibration blank solution and a mid-range calibration solution after each five to ten test solutions in order to monitor for baseline drift and sensitivity change, respectively.

11.3.3.3.4 Place the working calibration solution (11.3.3.2.1), the working calibration blank solution (11.3.3.2.2), the matrix modifier (7.6), and the unmodified sample and blank solutions in separate acid-washed autosampler cups and position as appropriate in the autosampler carousel. Analyse the matrix-modified calibration and test solutions, using the microprocessor or personal computer software to generate a calibration. Obtain a direct readout of sample and blank results in nanograms of lead per millilitre.

11.3.3.3.5 If significant baseline drift is observed during the course of analysis, or if the sensitivity changes by more than ±5 %, take one or more of the following corrective measures: either use the available software facilities of the microprocessor or personal computer to correct for sensitivity change ("reslope" facility); or suspend analysis and recalibrate the spectrometer. In either case, reanalyse the solutions that were analysed during the period in which the sensitivity change occurred.

11.3.3.3.6 If concentrations of lead above the upper limit of the linear calibration range are found, dilute the sample test solutions in order to bring them within the range of the calibration, and repeat the analysis. Make all dilutions so that the final nitric acid concentration is 1 + 1 or 1 + 9, as appropriate,

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and record the dilution factor (*DF*). Alternatively, analyse a smaller aliquot of sample, and make a correction for the amount of sample that is analysed.

11.3.3.3.7 Calculate the mean concentration of lead in the blank test solutions.

11.4 Estimation of the instrumental detection limit

11.4.1 Estimate the instrumental detection limit under the working analytical conditions following the procedure described below and repeat this exercise whenever the experimental conditions are changed.

11.4.2 Prepare test solutions at a concentration of 0,1 µg/ml of lead for flame atomic absorption analysis or 1 ng/ml for electrothermal atomic absorption analysis by diluting the working lead standard solution (7.8). Make these dilutions so that the final nitric acid concentration is 1 + 9.

11.4.3 Analyse the test solution at least ten times under repeatability conditions and calculate the instrumental detection limit as three times the sample standard deviation of the mean concentration value (see for example ISO 6955).

NOTE The limit of detection calculated from results using this procedure is an instrumental detection limit. This is of use in identifying changes in instrument performance, but it is not a method detection limit[7]. The instrumental detection limit is likely to be unrealistically low because it only takes into account the variability between individual instrumental readings; determinations made on one solution do not take into consideration contributions to variability from the matrix or sample.

11.5 Estimation of the method detection limit and method quantification limit

11.5.1 Estimate the method detection limit and method quantification limit under the working analytical conditions following the procedure described in 11.5.2 through 11.5.3 and repeat this exercise whenever the experimental conditions are changed significantly.

11.5.2 Prepare at least ten test solutions from laboratory blanks, following the sample preparation procedure in 11.2, and analyse the test solutions for lead under repeatability conditions.

If there is no measurable response from the instrument, fortify at least ten sampling substrates (8.2) with lead near the anticipated detection limit, for example, 1 µg of lead for flame atomic absorption analysis or 0,01 µg of lead for electrothermal atomic absorption analysis, by spiking the sampling substrate with 0,1 ml of a suitable calibration solution (7.8) diluted by an appropriate factor with 1 + 9 nitric acid (7.4). Prepare test solutions from the fortified filters, following the sample preparation procedure in 11.2, and analyse the test solutions for lead under repeatability conditions.

11.5.3 Make atomic absorption measurements on the test solutions derived from each test solution (11.5.2) (after carrying out digestion of the filters) and calculate the method detection limit and method quantification limit as three times and ten times the sample standard deviation of the mean concentration value, respectively.

NOTE An alternative procedure for estimating the method detection limit involves the analysis of samples fortified with the analyte of interest at values spanning the predicted detection limit[12].

11.6 Quality control

11.6.1 General

Quality control (QC) samples to process with each batch of field samples are summarized below.

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11.6.2 Reagent blanks and media blanks

Carry reagent blanks (water and reagents) and media blanks (unspiked sampling substrate) throughout the entire sample preparation and analytical process to determine whether the samples are being contaminated from laboratory activities. Process reagent blanks according to a frequency of at least one per 20 samples or a minimum of one per batch.

11.6.3 Spiked samples and spiked duplicate samples

11.6.3.1 Process these samples on a routine basis to estimate the method accuracy on the sample batch, expressed as a percent recovery relative to the true spiked value. Spiked samples and spiked duplicate samples consist of sampling substrate to which known amounts of analyte were added. (This can be accomplished by spiking known volumes of known concentrations of lead solution at amounts within the dynamic range of the instrument. The lead solution used shall be prepared from a stock standard solution from a different source than that used for preparing the calibration solutions.) Process these QC samples at a frequency of at least 1 per 20 samples or minimum of one per batch.

11.6.3.2 Monitor the performance of the method by plotting control charts of the relative percent recoveries and of the relative percent differences between the spiked samples and the spiked duplicate samples. If QC results indicate that the method is out of control, investigate the reasons for this, take corrective action and reanalyse the samples if necessary. See ASTM E882[15] for general guidance on the use of quality control charts.

11.6.4 Certified reference materials

Certified reference materials (CRMs) for lead, prepared in accordance with ISO 17034, shall be analysed prior to routine use of the method, and periodically thereafter, to establish that the percent recovery relative to the certified value is satisfactory. Suitable CRMs are available from many sources. A minimum of one CRM sample shall be analysed at least six times quarterly.

11.6.5 External quality assessment

If laboratories carry out lead in air analysis on a regular basis, it is strongly recommended that they participate in a relevant external quality assessment scheme or proficiency testing scheme.

12 Expression of results

12.1 Calculation

Calculate the mass concentration of lead in the air sample, c_{Pb}, in milligrams per cubic metre, at ambient conditions, using Formula (1):

$$c_{Pb} = \frac{\left(c_{\mathrm{Pb},1} \times V_1 \times DF\right) - \left(c_{\mathrm{Pb},0} \times V_0\right)}{V} \tag{1}$$

where

$c_{\mathrm{Pb},0}$ is the mean lead concentration, in micrograms per millilitre, in the blank test solutions;

$c_{\mathrm{Pb},1}$ is the lead concentration, in micrograms per millilitre, in the sample test solution;

V is the volume, in litres, of the air sample;

V_0 is the volume, in millilitres, of the blank test solution;

V_1 is the volume, in millilitres, of the sample test solution;

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DF is the dilution factor (*DF* = 1 in the absence of dilution).

12.2 Method performance

12.2.1 Sample collection

A collection efficiency of 1,00 was determined for the filter collection step for laboratory-generated lead nitrate aerosols and for lead fume[16].

12.2.2 Hot plate digestion and flame atomic absorption spectrometry

12.2.2.1 The detection limit of flame atomic absorption measurements depends in part on the instrument used. However, the detection limit of the method has been estimated to be 0,25 µg per sample, and the precision of the measurement procedure was <0,1 for samples in the range 0,9 µg to 2,25 µg and <0,03 for samples in the range 3,6 µg to 288 µg[6]. No bias has been identified. The applicable range is 1 µg to 200 µg Pb per sample, without dilution.

12.2.2.2 In tests using laboratory-generated lead nitrate aerosols[15], the coefficient of variation for a similar procedure was found to be 0,072 for the overall sampling and analytical method in the range 130 µg to 400 µg Pb/m^3. The bias of the method was found to be insignificant. Also, data from interlaboratory proficiency testing for samples of paint, soil and dust[16], [17] have indicated insignificant bias.

NOTE If the sample dissolution procedure using concentrated nitric acid and hydrogen peroxide is ineffective for the dissolution of particulate lead compounds present in the test atmosphere (for example, lead silicates), and an alternative, more vigorous dissolution procedure has not been used (for example, employing hydrofluoric acid), then the analytical results will be subject to a negative bias. If it is desired to determine lead in samples containing high concentrations of silicates, consider the use of hydrofluoric acid in the digestion procedure.

12.2.3 Microwave assisted digestion and flame atomic absorption spectrometry

Interlaboratory evaluations of lead determinations in reference materials (paint, soil and dust) have shown that microwave assisted digestion with concentrated nitric acid performs equivalently to hot-plate digestion when followed by lead analysis using flame atomic absorption spectrometry[18].

12.2.4 Ultrasonic extraction and flame atomic absorption spectrometry

Ultrasonic extraction with electrochemical determination of lead has been shown to perform equivalently to microwave assisted digestion and flame atomic absorption spectrometry in the determination of lead from laboratory-generated lead fume atmospheres[19].

12.2.5 Hot plate digestion and electrothermal atomic absorption spectrometry

The detection limit of electrothermal atomic absorption measurements depends in part on the instrument used. However, the detection limit of the method has been estimated to be 0,003 µg per sample, and the precision of the measurement procedure was < 0,05 for samples in the range 0,1 µg to 4,5 µg. No bias has been identified. The applicable range is 0,01 µg to 0,5 µg Pb per sample[6], without dilution.

12.2.6 Microwave assisted digestion and electrothermal atomic absorption spectrometry

Interlaboratory analysis of lead in reference materials has demonstrated the equivalence of microwave assisted digestion followed by electrothermal atomic absorption spectrometry to hot-plate digestion followed by electrothermal atomic absorption spectrometric determination of lead[18].

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12.2.7 Ultrasonic extraction and electrothermal atomic absorption spectrometry

Ultrasonic extraction with electrochemical detection of lead was evaluated against hot-plate extraction and electrothermal atomic absorption spectrometry for air samples collected from construction sites.[20] The test procedure using ultrasonic extraction was found to be equivalent to that using hot-plate digestion.

13 Special cases

13.1 If there is any doubt as to the suitability of the digestion or extraction procedure used for the dissolution of particulate lead compounds (for example, silicates) that can be present in the test atmosphere, determine its effectiveness by analysing a bulk sample of known lead content that is similar in nature to the material being used or produced. If the efficiency of recovery is less than 90 %, use an alternative, more vigorous dissolution procedure, for example, by using hydrofluoric acid. Do not use a correction factor to compensate for an apparently ineffective dissolution procedure.

It should be recognized that the recovery of lead can be dependent upon the particle size distribution of a bulk sample.

13.2 Anions that give rise to precipitates can interfere with lead analysis. If such interferents are likely to be present in sample solutions, add the disodium salt of ethylenediamine tetraacetic acid (EDTA) to the sample and blank solutions and to the calibration standard solutions, such that these solutions have a concentration of 0,1 mol/l of EDTA.

The addition of EDTA usually prevents precipitation, but high levels of phosphate can diminish the lead signal even in the presence of EDTA. If high levels of phosphate are suspected in the sample solutions, then the method of standard addition should be used to obtain accurate results (see ISO 6955).

13.3 It has been postulated[19], [21] that gaseous lead can be present in significant concentrations in certain work environments, for example, when high temperature processes are used. In such circumstances, the sampling method described in this document can possibly be less than fully effective because gaseous lead can pass through the sampling substrate. If necessary, this possibility can be investigated using a sampling train consisting of a filter and bubbler[19], [21].

13.4 When the transport cassettes or samplers are opened, it is advisable to look for evidence that particles have become dislodged from the sampling substrate during transportation. If this appears to have occurred, consider whether to discard the sample as invalid, or whether to wash the internal surfaces of the transport cassette or sampler into the sample dissolution vessel in order to recover the material concerned.

13.5 The load on the graphite tube or the graphite platform in the atomic absorption spectrometry is strongly dependent on the matrix of the samples to be analysed. For the specified temperature/time programme, the stability of the graphite furnace and thus the constancy of the recovery for samples with low arsenic content is about 24 hours.

14 Test report

The test report shall contain the required elements for test reports identified in ISO/IEC 17025, and also the following information:

— statement to indicate the confidentiality of the information supplied, if appropriate;

— complete identification of the air sample, including the date of sampling, the place of sampling, the type of sample (personal or static), either the identity of the individual whose breathing zone was sampled (or other personal identifier) or the location at which the general occupational environment

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was sampled (for a static sample), a brief description of the work activities that were carried out during the sampling period, and a unique sample identification code;

— reference to this document;

— make, type and diameter of sample substrate used;

— make and type of sampler used;

— make and type of sampling pump used, and its identification;

— make and type of flowmeter used, the primary standard against which the calibration of the flowmeter was checked, the range of flow rates over which the calibration of the flowmeter was checked, and the atmospheric temperature and pressure at which the calibration of the flowmeter was checked, if appropriate (see 10.1.3);

— time at the start and at the end of the sampling period, and the duration of the sampling period, in minutes;

— mean flow rate during the sampling period, in litres per minute;

— mean atmospheric temperature and pressure during the sampling period, if appropriate (see 10.1.3);

— volume of air sampled, in litres, at ambient conditions;

— name of the person who collected the sample;

— time-weighted average mass concentration of lead found in the air sample (in mg/m^3), at ambient temperature and pressure, or, if appropriate, adjusted to reference conditions;

— analytical variables used to calculate the result, including the concentrations of lead in the sample and blank solutions, the volumes of the sample and blank solutions, and the dilution factor, if applicable;

NOTE If the necessary data (for example, the volume of air sampled) are not available to the laboratory for the above calculations to be carried out, the laboratory report can contain the analytical result in micrograms of lead per sample.

— type(s) of instrument(s) used for sample preparation and analysis, and unique identifiers(s);

— estimated detection limit and method quantification limit under the working analytical conditions;

— any operation not specified in this International Standard, or regarded as optional;

— name of the analyst(s) [or other unique identifier(s)];

— date of the analysis;

— any inadvertent deviations, unusual occurrences, or other notable observations.

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Annex A
(informative)

Guidance on filter selection

A.1 General

The following guidance is intended to help the user choose the most suitable filter for a particular application. It is not an exhaustive treatise on the subject, and covers only the basics of those matters that merit consideration. In many instances, similar considerations apply to the selection of other sampling substrates, such as polyurethane foams.

A.2 Collection efficiency

A.2.1 Most filters that are typically used for sampling airborne particulate matter have the required collection efficiency (see 8.2) for sampling both the respirable and the inhalable fractions of airborne particles. Such filters include depth filters, for example, glass or quartz fibre filters, and membrane filters, for example, mixed cellulose ester membrane filters and membrane filters made from polymers such as polyvinyl chloride (PVC) or polytetrafluoroethylene (PTFE).

A.2.2 Cellulose (paper) filters can have a collection efficiency below 99 % and are generally unsuitable for sampling airborne particles containing lead.

A.2.3 Certain processes carried out at elevated temperatures can produce ultrafine particles condensed from the vapour phase, known as fume. Filters used to sample airborne particulate matter can have a reduced collection efficiency for these very small particles, which are significantly less than 1 µm in aerodynamic diameter. However, the particles usually agglomerate soon after formation to produce larger particles that are efficiently collected. In general, filters that have a collection efficiency that meets the specification given in 8.2 are therefore suitable for sampling fume.

It has been shown in Kelvin et al , Journal of Occupational and Environmental Hygiene, 18:12, 555-569, DOI: 10.1080/15459624.2021.1985726, that the substrate used can impact the sample efficiency when using open face samplers like the IOM sampler. It is assumed that particles bounce off the substrate. As it is not clear if the bounced particles are part of the internal wall deposits and there is no fully validated procedure to include wall deposits this should also be taken into account when choosing the sampling substrate.

A.3 Dust-loading capacity

A.3.1 Membrane filters are manufactured from a variety of polymeric materials by a number of different processes. In each case the result is a thin, flexible disc of microporous material, with well-defined pore size, pore structure, pore density, etc. Retention of particles takes place on the surface of membrane filters, which results in their having a relatively low dust-loading capacity in comparison with depth filters. If an excessive amount of dust is collected on a membrane filter, this can result in blockage of the pores, and failure of the sampling pump. In addition, sample can be lost from the filter during handling or in transport. Sampling times should therefore be kept reasonably short when sampling with membrane filters in dusty environments, or depth filters should be used.

A.3.2 Depth filters consist of fibres that have been pressed together to form an irregular three-dimensional mesh. Particles are not only retained at the surface, but also within the structure of

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the filter. This gives them a significantly higher dust-loading capacity than membrane filters. In this respect, depth filters are a better choice than membrane filters when sampling for long periods in dusty environments. However, depth filters tend to have a higher metal content than membrane filters, and this needs to be considered when selecting the filter to be used if metals other than lead are to be determined.

A.4 Lead content

A.4.1 The lead content of the filters should be as low as possible, since it can make a significant contribution to the blank, the variability of which determines (in part) the lower limit of the working range of the analytical method. Exactly how low the lead content of the filters depends upon the applicable limit value. The lower limit of the working range of the analytical method should be less than the amount of lead that would be collected when sampling air at 0,1 times the limit value over the selected sampling period at the selected flow rate.

A.4.2 Membrane filters generally have a very low lead content, and in this respect are suitable for nearly all applications.

A.4.3 Glass fibre filters are unsuitable for use when measuring certain metals for which they have a high blank value. This is also true of quartz fibre filters, but to a lesser extent. Glass and quartz fibre filters do not present problems for lead measurements, but their potential use needs consideration if metals other than lead are to be measured.

A.5 Mass stability

A.5.1 If the filters are to be weighed in order to determine the amount of dust collected, it is important that they are reasonably resistant to moisture retention, so that blank mass changes that can occur as a result of changes in atmospheric conditions (temperature, humidity) are as low and as repeatable as possible.

A.5.2 If glass or quartz fibre filters are used, it is important that these are not excessively friable, since this can introduce weighing errors due to loss of filter material. Quartz fibre filters can be more friable than glass fibre filters. However, this disadvantage is counterbalanced by their lower metal content.

A.6 Solubility

A.6.1 The filters should be either wholly soluble or wholly insoluble using the selected sample preparation method. Partially dissolved filters can make subsequent handling of either the sample solutions difficult, or they can cause analytical error because of a matrix mismatch between sample solutions and calibration solutions. If PVC filters are chosen for sampling, it is recommended that microwave assisted, or ultrasonic dissolution procedures are used.

A.6.2 If the sample preparation method selected involves quantitative transfer of the sample solution to volumetric glassware prior to analysis, the filters used for sampling should preferably be soluble using the sample preparation method concerned. This reduces the chance of incomplete transfer of the sample solution.

A.6.3 Mixed cellulose ester membrane filters of 0,8 µm mean pore diameter are soluble in nitric acid, and these are suitable for use when this acid is used in the selected sample preparation environment. Quartz fibre filters are soluble in hydrofluoric acid and are suitable for use when this acid is used. Other filters can be equally suitable. Certain types of filters, such as PVC, may be partially soluble, and may cause problems for analysis.

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A.6.4 If sample solutions are to be made to volume in the sample dissolution vessel (for example, a graduated centrifuge tube), it is unimportant whether or not the filters are soluble using the selected sample preparation procedure.

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Annex B
(informative)

Sampler wall deposits

B.1 General

This annex provides information on the necessity for accounting for deposits of particulate matter on the interior walls of a transport cassette or sampler. Additional details are provided in ASTM D8358[22].

B.2 Samplers

Samplers for aerosols typically consist of a filter supported in a holder, though other collection substrates are also used, for example, impaction plates and foams. The entire device is considered to be an aerosol sampler. The sampling efficiency of an aerosol sampler is considered to be the air concentration calculated from the particles collected by the sampler compared to their concentration in undisturbed air. All aerosol samplers exhibit a decrease in sampling efficiency with increasing particulate aerodynamic diameter. Size-selective samplers are designed for a specific sampling efficiency over a range of aerodynamic diameters, known as a sampling convention (see ISO 7708), and the sampling efficiency of the sampler is considered with reference to the relevant sampling convention. In some sampler designs, for example, cyclones, there is an internal separator to achieve the required size selection.

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B.3 Collection efficiency

The collection efficiency of an aerosol sampler has four components:

— the aspiration efficiency;

— the transfer efficiency within the sampler (either from sampler inlet to the collection substrate or, if an internal separator is present, both from the sampler inlet to the internal separator and from the internal separator to the collection substrate);

— the penetration efficiency (through the internal separator, if present);

— the capture efficiency of the collection substrate (for example, filtration efficiency, when the collection substrate is a filter).

For any given sampler design, the various components depend on the particle aerodynamic size and air flow rate through the sampler. The aspiration efficiency also depends on wind speed and direction, while the sampler's angle to the vertical influences both aspiration and transport efficiency. Part of the sample will deposit on internal surfaces of the sampler as a result of losses during passage within the sampler. In addition, if the sampler is transported after sampling, particles deposited on the substrate can become dislodged and add to deposits already on the internal surfaces of the sampler (although this is likely of lesser importance, except when the collection substrate is overloaded with sample). If the design specification for the sampler is to include all aspirated particles, these losses need to be taken into account. Several studies have reported median values of Pb deposits on the walls for two commercially available samplers in common use (see Annex A of ISO 15202-1:2020)[23] that range from almost equal to 20 % to >50 %. No pattern can be discerned from these data that would allow the use of correction factors. For some samplers, the sample deposited on the collection substrate is considered to be the entire sample, i.e. wall deposits are not considered to be part of the sample. For other samplers, it is recommended that the wall deposits are evaluated. Other relevant information regarding particle

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deposits on the interior walls of sampler and particle losses during the transportation of samples is available in ISO 15202-1:2020[23].

B.4 Contribution to the uncertainty budget

Where an air sampling and analytical method includes a specific procedure for recovering and analysing wall deposits, this needs to be taken into account when estimating the expanded uncertainty of the method.

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Annex C
(normative)

Temperature and pressure correction

C.1 Temperature and pressure correction for the indicated volumetric flowrate

C.1.1 Bubble flowmeters are preferred for measuring the volumetric flowrate because the readings they give are independent of temperature and pressure. For other flowmeters, it can be necessary to apply a correction to the indicated volumetric flow rate if the temperature and pressure at the time of measurement is different to when the calibration of the flowmeter was checked.

C.1.2 A typical example of the need for a temperature and pressure correction is when a constant pressure drop, variable area, flowmeter is used to measure the volumetric flowrate. Such a flowmeter typically has a scale that is calibrated to standard conditions as defined by the manufacturer. In this instance, use Formula (C.1) to calculate a corrected air sample volume:

$$V_{\text{corr}} = q_v \times t \times \sqrt{\frac{p_1 \times T_2}{p_2 \times T_1}} \tag{C.1}$$

where

V_{corr} is the corrected volume, in litres;

q_v is the mean flow rate, in litres per minute;

t is the sampling time, in minutes;

p_1 is the atmospheric pressure, in kilopascal, during calibration of the sampling pump;

p_2 is the mean atmospheric pressure, in kilopascal, during the sampling period;

T_1 is the temperature, in kelvin, during calibration of the sampling pump;

T_2 is the mean temperature, in kelvin, during the sampling period.

Any other flowmeter can also require a correction for variation in pressure and temperature; follow the manufacturer's instructions for such corrections.

C.2 Recalculation of lead in air concentrations to reference conditions

If necessary (see 10.1.3.1), recalculate lead in air concentrations to reference conditions (for example, 273 K and 101,3 kPa), Formula (C.2):

$$p_{\text{Pb,corr}} = p_{\text{Pb}} \times \frac{101{,}3 \times T_2}{p_2 \times 273} \tag{C.2}$$

where

$p_{\text{Pb,corr}}$ is the concentration of lead in the air sample, in micrograms per cubic metre, at reference conditions;

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p_{Pb} is the concentration of lead in the air sample, in micrograms per cubic metre, at ambient conditions;

T_2 is the mean temperature, in kelvin, during the sampling period;

p_2 is the mean atmospheric pressure, in kilopascal, during the sampling period;

273 is the reference temperature, in kelvin;

101,3 is the reference atmospheric pressure, in kilopascal.

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